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6	MC1R polymorphism associated to plumage color variations in Coturnix chinensis.
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27	The melanocortin 1 receptor (MC1R) gene was investigated as a candidate for plumage
28	variations in Chinese painted quail, Coturnix chinensis. Four silent and two missense
29	nucleotide polymorphisms were identified. The correspondent amino acid changes,
30	p.Glu92Lys and p.Pro292Leu were found in <i>Blue Face</i> and <i>Red Breasted</i> animals,
31	respectively. Blue Face is a melanic phenotype similar to the co dominant Extended
32	Brown of Japanese quail, and both share the p.Glu92Lys mutation. The association of
33	p.Pro292Leu with the recessive Red Breasted was confirmed in 23 animals from an
34	experimental F2 cross.
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40	KEYWORDS
41	Quail, MC1R, plumage, color variation.
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The melanocyte production of black eumelanins and red phaeomelanins is mainly 51 52 regulated by the activity of the melanocortin 1 receptor (MC1R), and has great influence in fur or plumage color. The MC1R gene has only one exon and has been reported to 53 cause color variation in both mammals and birds (Andersson 2003). In chicken (Kerje et 54 al. 2003), quail (Nadeau et al. 2006) and bananaquit (Theron et al. 2001) the same 55 polymorphism, p.Glu92Lys, causes dominant black plumages likely through a MC1R 56 57 constitutive activation (Robbins et al. 1993). In contrast, recessive pheomelanic colors have been linked to the inactivation of the receptor, at least in mammals (Kijas et al. 58 1998). 59 60 Chinese painted quail (Coturnix chinensis) is the smallest of the domesticated quail species and has an accentuated sexual dimorphism in plumage color. While Wild 61 females display a brown coloration similar to that of Japanese quail, Wild males show a 62 63 general blue pattern on its back and a red tail (Figure 1). Because breeders value the colors of these animals, several plumage varieties have been already established. Some 64 of these are compatible with variations in melanins content, such as the incomplete 65 dominant Blue Face and maybe the recessive Red Breasted. Interestingly, although Blue 66 67 Face males and females have darkened eumelanic plumages, there is a strong contrast 68 among the *Red Brested* sexes: females display lighter brown colors while males display a completely blackened face and a ventral extended red patch (Figures 1). 69 70 71

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- 75 Thirteen quails of two different origins displaying *Wild* (5), *Blue Face* (3) and *Red*
- 76 Breasted (5) plumages were sampled to assess MC1R variability. A Wild male and a
- 77 *Red Breasted* female were used to generate an experimental cross and 15 *Wild* and 8
- 78 *Red Breasted* F2 individuals were analyzed. In all cases, DNA was extracted from
- 79 growing feathers as described in Vidal et al. (2010a).
- 80 Primers MC1R-F1 5'- ACGGCCCCAGCCAGGGGTCCT-3' and MC1R-R1 5'-
- 81 AGGCACACATCACTGCAAAG-3' were designed from published sequences of
- 82 Coturnix chinensis (AB201632) and Gallus domesticus (D78272.1) to amplify 1200 bp
- including the full coding region of the gene. Two internal primers (MC1R-F2 5'-
- 84 CCTCATCCTCATCGTCACCT-3' and MC1R-R2 5'-
- 85 TACCAGGAGCACAGCACCAC-3') were designed to fully sequence the 942 bp of
- the coding region of the *MC1R* gene. PCR products were purified with the ExoSAP-IT
- 87 PCR Product Cleanup (Thermo Fisher Scientific) and sequenced with the BigDye
- 88 Terminator v3.1 Cycle Sequencing kit (Thermo Fisher Scientific). Sequences were
- aligned using the Multalin software (Corpet 1988). The effects of new non-silent
- 90 polymorphisms were assessed with Panther-Psep (Tang & Thomas 2016).

- 92 The sequencing of 13 individuals allowed the identification of 6 polymorphisms
- conforming 5 haplotypes (see Table 1). Two of the polymorphisms are missense, and
- cause the amino acid changes p.Glu92Lys (haplotype H3) and p.Pro292Leu (haplotypes
- 95 H4 and H5).
- 96 The p.Glu92Lys change has been demonstrated to cause dominant melanism in mice
- 97 (Robbins et al. 1993), chicken (Takeuchi et al. 1996) and bananaquit (Theron et al.
- 2001). This melanism is related to a constitutive activation of the MC1R receptor,

99 which increases the concentration of cAMP inside the melanocytes, and eumelanins are

100 synthesized almost exclusively (Robbins et al. 1993; Ling et al. 2003).

101 Interestingly, in Japanese quail, this same polymorphism has been associated to the

102 darker *Extended Brown* plumage (Nadeau et al. 2006), an autosomal incomplete

103 dominant mutation. The constitutive activation of MC1R has been suggested as the

104 molecular mechanism for this phenotype as well (Nadeau et al. 2006).

105 We have found that all *Blue Face* animals carried one copy of haplotype H3, while none

106 of the others animals did. Because the pattern of inheritance of this melanic plumage

107 mimics the *Extended Brown* of Japanese quail, and although the number of available

samples is low, we suggest that an activation of MC1R caused by p.Glu92Lys is a likely

109 mechanism explaining this phenotype in Chinese painted quail.

110 The sequencing of 5 *Red Breasted* animals showed that three of them were homozygote

111 H4 and two were homozygote H5. These haplotypes differ in one silent position,

112 g.C601T, and both carry the g.C872T missense polymorphism causing a p.Pro292Leu

amino acid change (see Table 1). An experimental F2 cross was generated to confirm its

association with the *Red Breasted* plumage, and 15 *Wild* and 8 *Red Breasted* F2 animals

115 were genotyped. As expected all the *Red Breasted* animals were homozygote H4, while

of the *Wild* animals 10 were heterozygote H1H4 and 5 were homozygote H1.

117 The analysis of the p.Pro292Leu polymorphism using Panther-Psep (Tang & Thomas

118 2016) suggests a high probability of deleterious effects. So far, deleterious mutations of

this gene in mammals have been related to an increase in phaeomelanin production

120 caused by the receptor's inactivation. This mechanism has been described in yellow

mice (Robbins et al. 1993) and it is likely to explain red coloration in other mammals,

such as pig (Kijas et al. 1998). In humans, MC1R mutations causing red headedness are

123 considered to act in this same way, through deleterious mutations. Interestingly, a

- 124 polymorphism with very intense effects, p.Asp294His, has been located in the
- transmembrane domain 7 (Lightner 2008) which is therefore suspected to be
- 126 functionally relevant. Both p.Asp294His and p.Pro292Leu are thus highly likely to be
- 127 deleterious by altering this transmembrane domain.
- 128 In this sense, a hypothetical loss-of-function caused by p.Pro292Leu would be
- 129 compatible with *Red Breasted* being recessive; however, the colors found in *Red*
- 130 *Breasted* (especially males) do not match other typical *MC1R* phenotypes. In fact, clear
- 131 *MCIR* phaeomelanic phenotypes have not been found in birds yet (Roulin & Ducrest
- 132 2013), and brown and red plumages have been associated to *Sox10* in chickens
- (Gunnarsson et al. 2011) and pigeons (Domyan et al. 2014).
- 134 Interestingly enough, *MC1R* polymorphisms that could be inactivating the receptor have
- been linked to recessive dark phenotypes in guinea fowl (Vidal et al. 2010b) and turkey
- 136 (Vidal et al. 2010a). These phenotypes do not display the typical white dots found in
- 137 *Wild* guinea fowl nor the white bars of *Wild* plumage turkeys, thus resulting in a darker
- 138 general appearance. It could be possible then that these colorations are not related to
- 139 neat increases in eumelanin synthesis.
- 140 In this context, the association of the putatively deleterious p.Pro292Leu mutation to the
- 141 *Red Breasted* quail could be in consonance with the inactivation of the receptor in birds
- 142 not directly affecting eumelanin/phaeomelanin balance, but plumage pattern and/or
- color distribution. This could imply a significant difference in the function of MC1R of
- 144 mammals and birds that should be further studied.

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151	REFERENCES
152	Andersson, L. (2003). Melanocortin receptor variants with phenotypic effects in horse,
153	pig, and chicken. Annals of the New York Academy of Sciences 994, 313-8.
154	Baiao, P.C., Schreiber, E. & Parker, P.G. (2007). The genetic basis of the plumage
155	polymorphism in red-footed boobies (Sula sula): a melanocortin-1 receptor
156	(MC1R) analysis. Journal of Heredity 98, 287–292.
157	Corpet, F. (1988). Multiple sequence alignment with hierarchical clustering. Nucleic
158	Acids Research 16, 10881–90.
159	Domyan, E.T., Guernsey, M.W., Kronenberg, Z., Krishnan, S., Boissy, R.E., Vickrey,
160	A.I., Rodgers, C., Cassidy, P., Leachman, S.A., Fondon, J.W., Yandell, M. &
161	Shapiro, M.D. (2014). Epistatic and combinatorial effects of pigmentary gene
162	mutations in the domestic pigeon. Current Biology 24, 459–464.
163	Gunnarsson, U., Kerje, S., Bed'hom, B., Sahlqvist, A.S., Ekwall, O., Tixier-Boichard,
164	M., Kämpe, O. & Andersson, L. (2011). The Dark brown plumage color in
165	chickens is caused by an 8.3-kb deletion upstream of SOX10. Pigment Cell and
166	Melanoma Research 24, 268–274.
167	Kerje, S., Lind, J., Schütz, K., Jensen, P. & Andersson, L. (2003). Melanocortin 1-
168	receptor (MC1R) mutations are associated with plumage colour in chicken. Animal
169	Genetics 34, 241–248.
170	Kijas, J.M., Wales, R., Törnsten, A., Chardon, P., Moller, M. & Andersson, L. (1998).
171	Melanocortin receptor 1 (MC1R) mutations and coat color in pigs. Genetics 150,
172	1177–1185.

- Lightner, J.K. (2008). Genetics of Coat Color I: The Melanocortin 1 Receptor (MC1R).
  Answer Research Journal 108, 109–116.
- Ling, M.K., Lagerström, M.C., Fredriksson, R., Okimoto, R., Mundy, N.I., Takeuchi, S.
  & Schiöth, H.B. (2003). Association of feather colour with constitutively active
  melanocortin 1 receptors in chicken. European Journal of Biochemistry 270, 1441–
  1449.
- 179 Nadeau, N.J., Minvielle, F. & Mundy, N.I. (2006). Association of a Glu92Lys

substitution in MC1R with extended brown in Japanese quail (Coturnix japonica).

181 Animal Genetics 37, 287–289.

- 182 Robbins, L.S., Nadeau, J.H., Johnson, K.R., Kelly, M.A., Roselli-Rehfuss, L., Baack,
- E., Mountjoy, K.G. & Cone, R.D. (1993). Pigmentation phenotypes of variant
  extension locus alleles result from point mutations that alter MSH receptor
- 185 function. Cell 72, 827–834.
- 186 Roulin, A. & Ducrest, A.L. (2013). Genetics of colouration in birds. Seminars in Cell
  187 and Developmental Biology 24, 594-608.
- 188 Takeuchi, S., Suzuki, H., Yabuuchi, M. & Takahashi, S. (1996). Possible involvement
- 189 of melanocortin 1-receptor in regulating feather color pigmentation in the chicken.
- Biochimica et Biophysica Acta Gene Structure and Expression 1308, 164–168.
- 191 Tang, H. & Thomas, P.D. (2016). PANTHER-PSEP: predicting disease-causing genetic
- variants using position-specific evolutionary preservation. Bioinformatics 32,
  2230–2232.
- 194 Theron, E., Hawkins, K., Bermingham, E., Ricklefs, R.E. & Mundy, N.I. (2001). The
- 195 molecular basis of an avian plumage polymorphism in the wild: a melanocortin-1-
- receptor point mutation is perfectly associated with the melanic plumage morph of
- the bananaquit, Coereba flaveola. Current Biology 11, 550–557.

198	Vidal, O., Vinas, J. & Pla, C. (2010a). Variability of the melanocortin 1 receptor
199	(MC1R) gene explains the segregation of the bronze locus in turkey (Meleagris
200	gallopavo). Poultry Science 89, 1599–1602.
201	Vidal, O., Araguas, R., Fernández, R., Heras, S., Sanz, N. & Pla, C. (2010b). Melanism
202	in guinea fowl (Numida meleagris) is associated with a deletion of Phenylalanine-
203	256 in the MC1R gene. Animal Genetics 41, 656–658.
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## 228 Table 1. MC1R variation in Chinese painted quail.

Hanlatura	Conhonk	Alelle	DNA (Amino acid) <sup>1</sup>					
паріотуре	Genbank		117	271 (92)	601	633	714	872 (292)
H1	MG520490	Wild	С	G (Glu)	С	G	G	C (Pro)
H2	MG520492	Wild	Т	-	-	A	-	-
H3	MG520491	Blue Face	-	A (Lys)	-	-	A	-
H4	MG520493	Red Breasted	-	-	-	-	-	T (Leu)
H5	MG520494	Red Breasted	-	-	Т	-	-	T (Leu)

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-	-	-

<sup>1</sup>Position in Japanese quail protein sequence BAD91489.1.



Figure 1. Plumages of *Wild* and *Red Breasted* quail. (A) *Wild* female, (B) *Red Breasted* female, (C) *Red Breasted* –front- and *Wild* –back- males. A complete color chart can be found at http://www.featheredobsessions.com/Button\_Quail\_Colors.php.