

ABSTRACT

 This study evaluates the link between sea warming and three important aspects of the population dynamics of bluefish, *Pomatomus saltatrix*, in the NW Mediterranean: condition (health measures), reproduction and abundance (as reflected in catch per unit effort, CPUE). We compared the bluefish populations from two areas with different thermal regimes: the Gulf of Roses and the Ebre Delta. First, in order to compare the condition and reproduction parameters of the two populations, specimens were collected from the commercial catch landed in two ports located in these areas. Several indicators of fish condition and reproduction such as hepatosomatic index, gonadosomatic index, maturation timing, length-at-first maturity, fecundity, egg quality and atresia (ovarian follicle resorption) were analysed and compared between study areas. Secondly, time series analyses of sea surface temperatures (SST) and monthly bluefish CPUEs in the two study areas were assessed using Box-Jenkins transfer function models. Our results indicate that bluefish benefit from sea warming in the NW Mediterranean, particularly in the northernmost zone, as evidenced by increasing fish condition in these waters. Sea warming may favour the changes in spawning phenology and the northward expansion of this species. This study provides evidence that a marine fish species that is able to expand or it is forced to shift its distribution range due to sea warming, may take advantage of other environmental conditions of new habitats, contributing to the enhancement of the population's productivity in these areas.

 KEYWORDS: NW Mediterranean, sea surface temperature, *Pomatomus saltatrix*, fish condition, reproductive biology, predator.

INTRODUCTION

 The Mediterranean Sea is a semi-enclosed basin where sea warming is likely to be more pronounced and to become apparent sooner than in other more open oceans (Astraldi *et al.*, 1995; Bethoux *et al.*, 1999; Walther *et al.*, 2002; Coll *et al.*, 2010). Climate models predict that the Mediterranean basin will be one of the regions that is most affected by the ongoing warming trend and by an increase in extreme events (Parry, 2000; Lejeusne *et al.*, 2010; Calvo *et al.*, 2011; Durrieu de Madron *et al.*, 2011). Subtropical species are found mainly in the eastern basin and southern Mediterranean, where water temperatures are higher, whereas cold-temperate species inhabit the cooler northern areas (Salat, 1996; Bianchi and Morri, 2000; Coll *et al.*, 2010).

 A direct consequence of sea warming is a simultaneous increase in the abundance of thermo- tolerant species and the decline or disappearance of 'cold' stenothermal species (Lejeusne *et al.*, 2010). In a semi-closed system such as the Mediterranean basin these changes may cause the progressive homogenization of the marine biota, the dislocation of other species, the loss of endemic species coupled with invasion by non-indigenous species and/or cascade effects on food webs (Molinero *et al.*, 2005).

 Not only could marine biodiversity be seriously affected, but climate change may also affect fisheries directly by influencing fish stocks and the global supply of fish for consumption. Commercial fishing would also be affected by changes to the overall value of fish catches due to changes in the abundance of the species caught. The value of the total catch may decrease as warm water species - which can be of lower commercial value - increase at the expense of other species of greater commercial value, e.g. the increase of the thermophilic, low value *Sardinella aurita* (Sabatés *et al.*, 2006) vs. the decrease of the temperate, high value *Engraulis encrasicolus* and *Sardina pilchardus* (Martín *et al.*, 2011).

 The bluefish, *Pomatomus saltatrix*, (Linnaeus 1766), is distributed over continental shelves and in estuaries of temperate waters throughout most of the world, with the exception of the northern and mid-Pacific Ocean (Juanes *et al.*, 1996). Relatively little is known about the biology of this species in the Mediterranean. Sabatés and Martín (1993) provided a number of observations regarding its presence in certain areas, and its relative importance in terms of fishing activity as well as the duration of spawning. They proposed the southern Catalan as the northernmost boundary of bluefish distribution in the western Mediterranean Basin. However, more recently, Sabatés *et al.* (2012) found that the bluefish distribution range was expanding northwards due to the increasing water temperatures allowing reproduction to take place in more northern areas, as revealed by the presence of bluefish larvae in plankton. Furthermore, it has been suggested that shifts in the bluefish distribution range could be the result of current climate change and that correct management of fish communities required this issue to be assessed (Goodbred and Graves, 1996; Juanes *et al.*, 1996; Pardiñas *et al.*, 2010). This warm water species appears to be a good candidate for investigating the effects of sea warming.

 The aim of this study was to evaluate the influence of sea temperature on three important aspects of population dynamics of bluefish in the NW Mediterranean: condition, reproduction and abundance (as reflected in catch per unit effort). The hypothesis behind this study is that the abundance, condition and reproduction of bluefish in coastal waters off the north-western Mediterranean Sea are all being influenced by sea warming, manifested through changes in fish condition and spawning characteristics, which may have resulted in increased catches over time.

METHODS

Sampling locations

 Samples were collected from two areas with different thermal regimes in the Northwest Mediterranean Sea: the more northerly Gulf of Roses and the more southerly Ebre Delta (Fig. 1). The Gulf of Roses and surrounding waters have lower mean water temperatures than the Ebre Delta (Salat *et al.*, 2002).

Fish condition and reproduction evaluation

 In order to compare the condition and reproduction parameters of the bluefish between the two study areas, samples of bluefish were collected monthly from the commercial catch landed at the port of Roses (Gulf of Roses) from July 2010 to August 2012 and at the port of Sant Carles de la Ràpita (Ebre Delta) from May to August 2012, shortly after the small-scale and bottom-trawl fishing vessels had landed their catches. In the laboratory, we recorded the total length (TL) to the nearest 0.5 cm and total weight (TW) to the nearest 1 mg. Subsequently, the bluefish specimens were dissected and eviscerated, and the somatic or eviscerated body weight (SW), the liver weight (LW) and the gonad weight (GW) were obtained to the nearest 1 mg. Finally, the gonads were fixed in buffered formalin (4%) for further histological processing and fecundity estimation.

 The hepatosomatic (HSI) and gonadosomatic (GSI) indices were calculated for each individual. All indices were calculated based on SW in order to avoid possible variations arising from differences in the contents of the digestive tract. The hepatosomatic index (HSI) was calculated as HSI = 100 (LW/SW), where LW and SW represent liver and somatic wet weights, respectively. The liver is the main energy store of many fishes and therefore the HSI is expected to provide the best estimate of bluefish condition. Finally, the gonadosomatic index (GSI) was estimated using the formula GSI = 100(GW/SW), where GW and SW represent gonad and somatic wet weights, respectively.

 Although the sex and reproductive status of specimens were first macroscopically determined, a histological analysis was also performed in order to provide a more accurate analysis of the reproductive characteristics and the annual reproductive cycle of bluefish. Thus, a histological study of the gonads of every single individual was carried out in order to determine the stages of development of their germ cells. Central portions (transverse sections) of the fixed gonads were dehydrated and embedded in paraffin, sectioned at between 3-8 µm, depending of their state of maturity, and stained with haematoxylin-eosin and Mallory's trichrome. The latter staining method

 highlights the *zona radiata* and its continuity and facilitates the detection of atretic oocytes (Muñoz *et al.*, 2010). Maturation stages of the gonads were classified in line with Brown-Peterson *et al.* (2011).

113 In order to define sexual maturation as a function of body length, the L_{50} (body length at which 50% of the individuals were mature) was estimated separately for samples taken from the Gulf of Roses and from the Ebre Delta. To predict the probability that an individual was mature based on its length, binary maturity observations (0=immature, 1=mature) and length (TL; cm) were fitted to binary logistic models to construct maturity ogives based on logistic equations based on 1 cm length-classes. Size at maturity was estimated from 75 and 103 females (length 26-91 cm TL) and from 63 and 84 males (length 26-79 cm TL) sampled at the Gulf of Roses and the Ebre Delta, respectively.

 The presence of hydrated oocytes and postovulatory follicles (POF) was determined to select suitable specimens for the analysis of fecundity. The oocyte size-frequency distribution for each spawning female was analysed in order to detect the batches of oocytes, since this may demonstrate different maturation patterns and associated fecundities (Kjesbu *et al.*, 1998). Thus, fecundity was estimated for each specimen in 'Spawning Capable' (SC) or 'Actively Spawning' (AS) phases using the Gravimetric Method combined with Image Analysis as explained by Murua *et al.* (2003). With this aim, subsamples of about 150 mg were taken from the central area of the ovary; oocytes were separated from connective tissue using a washing process (Lowerre-Barbieri and Barbieri, 1993) and sorted by size through several sieves (from 1000 to 100 µm), which facilitated the next steps of counting and measuring oocytes using a computer-aided image analysis system (Image-Pro Plus 5.1; [www.mediacy.com\)](http://www.mediacy.com/). In order to explore the size distribution for each oocyte developmental stage, the mean diameter of 200 oocytes from each stage were measured from the histological sections as the average of major and minor axes. Due to their irregular shape the mean diameter of the hydrated oocytes was estimated separately after adding glycerine, which facilitates their differentiation.

 Estimations of fecundity were based on Batch Fecundity (BF) defined as the number of eggs spawned per batch (Murua and Motos, 2006; Domínguez-Petit and Saborido-Rey, 2010). This was 138 estimated as $BF = GW*(O/Sw)$, where GW is the gonad weight after fixation, O is the number of hydrated oocytes in a weighted subsample of ovarian tissue, Sw is the subsample weight. The Relative Batch Fecundity (RBF) was also calculated, as the value of batch fecundity per gram of somatic weight of the fish.

 The dry mass and diameter of hydrated oocytes were used to estimate the quality of the oocytes, hence an approximation of the reproductive success (Brooks *et al.*, 1997). Thus, mean dry weights 144 were estimated by drying (for 24 h at 110°C) two replicates per sample of the oocytes.

145 The prevalence of atresia, P_a (percentage of sexually mature females that have α -atretic vitellogenic oocytes) and the relative intensity of atresia, *RIa* (percentage of α-atretic vitellogenic oocytes in relation to the total number of vitellogenic oocytes) was estimated from observations at three different focal planes of different histological slides of each specimen (Kurita *et al.*, 2003).

Statistical analyses

 Generalized linear models (GLMs) (McCullagh and Nelder, 1989) were used to investigate the variation of fish condition with sex, stage of maturity and size. GLMs were fitted to HSI and GSI as response variables and using as categorical predictor variables the sex (females and males) and maturity (immature, developing, spawning capable, regressing, regenerating) and as continuous predictor variables the size (total length). All predictors and their first order interactions were initially included in the GLM. Analysis of deviance to evaluate the significance (F-test) of the factors in the model was performed by a stepwise procedure, and the most appropriate error models were chosen on the basis of residual plots. GLMs incorporating the sex and maturity as predictor variables accounted significantly for 36.4% of the deviance of HSI and 43.4% of the variability in GSI (Table 1). Size did not significantly accounted for the deviance neither of HSI nor of GSI (p>0.05). Thus, we used the GLM approach to standardize HSI and GSI data only for the effects of

 sex and maturity by estimating the adjusted means of HSI and GSI as the means after adjusting for the variation of the covariables (sex and maturity).

 In order to carry out the comparative analyses between sampling areas, a sub-dataset of the samples from Gulf of Roses was created matching the temporal scale of the Ebre Delta (from May to August 2012) so that the spatial variation could be evaluated. This analysis indicated that there were significant differences between sampling areas in the condition and reproduction variables, e.g. in GSI (ANOVA, F1,238=4.67, p=0.032) or HSI (ANOVA, F1,238=4.91, p=0.028). Furthermore, there was an insignificant interannual difference (2010, 2011 and 2012) in the Gulf of Roses, e.g. 169 GSI (ANOVA, $F_{2,137}=2.05$, p=0.133) or HSI (ANOVA, $F_{2,137}=2.37$, p=0.097). All these analyses support the grouping of all three reproductive annual cycles of the Gulf of Roses in order to compare each variable to those of the Ebre Delta.

 Subsequently, for all aforementioned variables the Shapiro-Wilk test was used to test the assumptions of normality and Levene's test was used to test the homogeneity of variances (Zar, 1996). Once assumptions were checked and met, ANOVA models were used to compare all indices 175 between sampling areas. A p-value of α =0.05 or less was considered to be statistically significant. Moreover, if the ANOVA indicated significant differences Bonferroni's multiple tests were applied for post hoc comparisons of significant effects (Sokal and Rohlf, 1995). Conversely, if the assumptions were not met, the data were analysed with a non-parametric Mann-Whitney U test 179 (Sokal and Rohlf, 1995). A p-value of $\alpha = 0.05$ or less was considered to be statistically significant.

 Finally, the relationship between batch fecundity (BF) and the total length (TL) was estimated by fitting power functions. Then, ANOVAs (for relative batch fecundity), ANCOVAs (for batch fecundity using TL as covariate) and paired Student's tests (for oocyte quality and relative intensity of atresia) were also used to find out whether, on average, differences between sampling areas in the aforementioned variables were statistically significant (Zar, 1996).

 Trends in sea surface temperatures (SSTs) and bluefish landings in Catalan waters have already been explored by Sabatés *et al.* (2012), who found that both parameters showed a tendency to increase from 1997 to 2010 (particularly during spring months). They also explored the seasonality of bluefish landings and observed a seasonal pattern, closely matching that of the SSTs, in the monthly landings over the year in which landings were almost nil during the colder months but began increasing at the time SSTs also began to increase; subsequently, landings decreased as SSTs decreased. Thus, our analyses focuses on determining and quantifying the possible relationships between the monthly time series of bluefish catch per unit effort (CPUE) and SST using bivariate ARIMA (autoregressive-integrated-moving-average) transfer functions (Box and Jenkins, 1976).

 First, the data on sea surface temperature (SST, in °C) from both areas (the Gulf of Roses and the Ebre Delta) was retrieved from the International Comprehensive Ocean-Atmosphere Data Set (ICOADS) for the period 2000-2011 (ds540.0-Release 2.5) (Woodruff *et al.*, 2011). The data comprised individual daily mean sea surface temperature observations for 1° latitude x 1° longitude units. In this study, we used the time series from two 1° squares in the north-western Mediterranean: the Gulf of Roses (42.0 to 43.0°N and 3.0 to 4.0° E) and the Ebre Delta (40.0 to 41.0° N and 0.0 to 1.0° E). The mean monthly and seasonal values of SST in both areas were calculated by averaging daily temperatures retrieved from the ICOADS data. The seasons were defined as follows: winter (January, February, and March), spring (April, May, and June), summer (July, August, and September) and autumn (October, November, and December).

 Secondly, data was collected - on total monthly bluefish landings (recorded in kg) and nominal fishing effort (number of vessels), from the fishing ports along the Catalan coast (Fig. 1) - from the fishery statistics of the Department of Agriculture and Fisheries of the Government of Catalonia (2000-2011) [\(www20.gencat.cat/portal/site/DAR\)](http://www20.gencat.cat/portal/site/DAR). It should be noticed that bluefish landings included only those catches landed by the small-scale fleets (devoted to artisanal fishing and whose fishing grounds are located in the vicinity of the base port) because it is the more prevalent vessel type that catches most of the bluefish landings (from 90 to 95%) in all ports. The occasional landings recorded by the bottom trawler and purse-seine fleets were not included, which have been suggested previously and used by Sabatés *et al.* (2012). Since there has been a considerable reduction in the fishing effort in Catalan waters in recent years (Martín *et al.*, 2011; Catalonia Fisheries Statistics, 2012; Sabatés *et al.*, 2012), the monthly number of active vessels at each port was used as a proxy of fishing effort at each port. In this way, the time series of the monthly catches per unit effort (CPUE), expressed as kg per vessels, were estimated for different periods (2000- 2011) at two fishing ports from the Gulf of Roses (Roses and L'Escala) and three from the Ebre Delta area (L'Ametlla de Mar, L'Ampolla, and Sant Carles de la Ràpita).

 Finally, in order to determine and quantify the possible relationships between the time series of bluefish CPUEs and SST, bivariate ARIMA transfer function models were constructed using the statistical package ForeTESS (Prat *et al.*, 2010). These models describe the time-lagged relationship between the dependent (CPUE) and the independent (SST) variables while taking into account the autocorrelation of the disturbance of the data (Box and Jenkins, 1976).

 In order to look at the temperature thresholds related to the timing of the arrival of bluefish in both study areas as environmental conditions become favourable to this species, we repeated the transfer function models but this time using only the values of those months when SST increased by 229 a factor of 1°C, from 13°C (minimum SST observed) to 27°C (maximum SST observed): SST_{13} + 230 SST₁₄ + ... +SST₂₇, and only physical values from months when SST was higher than 13^oC were included in the transfer function models and the rest of the months when temperature was lower than 13°C were replaced by zeroes, which meant 'no influence on CPUE' in our models. In this way, several transfer function models were explored and applied to the 13-27°C SST range so that a minimum SST threshold could be found for each time series. Models were compared using Akaike's information criterion (AIC) to determine which model offered the best fit for SST thresholds at each time series.

RESULTS

 A total of 138 bluefish (75 females and 63 males) were sampled in the Gulf of Roses, and 187 (103 females and 84 males) in the Ebre Delta. The fish length ranged from 26.0 to 91.0 cm TL.

Fish condition and reproductive biology

 Since bluefish specimens were not significantly different between sampling areas in size 242 (ANOVA, $F_{1,324}=2.86$, p=0.092) or sexes (ANOVA, $F_{1,324}=3.24$, p=0.073), data were pooled (grouped into a single dataset per sampling area).

 HSI (standardized based on GLMs for sex and maturity) were significantly (ANOVA, 245 F_{1,324}=4.56, p=0.034) higher in the Gulf of Roses (1.56 \pm 0.48) than in the Ebre Delta (1.26 \pm 0.31) indicating better fish conditioning in the northern population (Fig. 2). Significant differences were 247 also found in GSI (ANOVA, $F_{1,324}=5.97$, p=0.015), in which case specimens sampled at the Gulf of 248 Roses showed higher GSI (3.64 ± 2.54) compared to specimens from the Ebre Delta (1.86 ± 1.39) indicating that the reproductive investment was much higher in the northern population (Fig. 2). In terms of the time of the year, in both sampling areas, the GSI was low during spring and then increased sharply, peaking in the summer before declining again in autumn (Fig. 3).

 Differences in the timing of gonadal development were also observed between sampling areas (Fig. 3). In females from the Gulf of Roses, the early stages of development were observed mainly during the spring (April to June); then, during the summer (July to September), spawning activity occurred; and subsequently, the rate of spawning activity declined in late summer (September) and autumn (October to December) when all the females were in the regeneration stage (Fig. 3a). Similarly, the spawning season of females from the Ebre Delta is mainly during the summer, although 'Spawning Capable' females were also observed earlier during the spring (which was not the case in the Gulf of Roses) (Fig. 3b). In addition, later on in the summer, the proportion of 'Regressing' females from the Ebre Delta (28.9%) was higher than it was in the Gulf of Roses in

 the same season (4.4%), indicating that not only did spawning activity begin earlier in the Ebre Delta but also that the rate of spawning activity there declined earlier than it did in the Gulf of Roses. Similar differences between the two sampling areas were observed in the timing of gonadal development of bluefish males (Fig. 3). In the Gulf of Roses the early stages of development and 'Spawning Capable' males were observed in the spring, but spawning activity did not peak until the summer, as revealed by the presence of specimens in the samples that were in 'Actively Spawning' and 'Regressing' stages. However, in the Ebre Delta, although the spawning season of males also takes place mainly in summer, some 'Actively Spawning' males were already observed in spring (Fig. 3d), suggesting that the spawning activity of males, as was the case with females, began earlier in the Ebre Delta than it did in the Gulf of Roses.

271 The mean length at which 50% specimens were mature (L_{50}) was, for females, 37.5 cm TL in the 272 Gulf of Roses and 37.7 cm TL in the Ebre Delta. For males, the L₅₀ in the Gulf of Roses was 36.5 cm TL while in the Ebre Delta it was 36.2 cm TL. In both sampling areas all females and males were mature at about 40 cm TL.

 The oocyte size-frequency distribution in bluefish females showed a continuous size-frequency development of oocytes except for ovaries in the 'Actively Spawning' phase, which had a separate mode of very large (>600 μm) hydrated oocytes (Fig. 4). Only when hydration occurred just before ovulation, did hydrating oocytes outgrow the standing stock of vitellogenic oocytes and a separate mode of mature hydrated oocytes developed. The presence of hydrated oocytes alongside the standing stock of developing oocytes in individual ovaries, as well as the presence of post-ovulatory follicles (POFs) in ovaries with advanced vitellogenic oocytes, indicates that asynchronic multiple batch spawning occurs in this species.

 The specific size ranges for each stage of secondary oocyte development are shown in Table 2. Significant differences (Mann-Whitney U test) in the median diameter of oocytes from the two 285 sampling areas were found at nearly all stages of development: at CA (U=4331.5, p<0.05), at Vtg-1

 (U=8459.5, p<0.05), at Vtg-2 (U=4488.5, p<0.05), at Vtg-3 (U=4224.5, p<0.05), and at GVM (U=4738.0, p<0.05), with Gulf of Roses oocytes being always slightly larger than Ebre Delta oocytes. However no significant differences were observed in the median diameters of hydrated oocytes (U=808.5, p=0.48).

 Only 13 females (5 from the Gulf of Roses and 8 from the Ebre Delta) met the histological criteria (actively spawning with hydrated oocytes and without POFs) for fecundity analysis. The batch fecundity (BF) of bluefish ranged from 84009 to 259636 eggs per spawning batch in fish ranging from 41.0 to 76.5 cm TL. The relationship between BF and TL was fitted to the following 294 exponential regression model: $BF=832.45 \cdot TL^{1.275}$ ($r^2=0.652$, n=13). Among sampling areas, the mean batch fecundities were estimated at 166559(±62062) and 153653(±42613) eggs per spawning batch for the Gulf of Roses and the Ebre Delta, respectively; and the mean relative batch fecundities 297 (RBF) were estimated at $102.1(\pm 32.5)$ and $92.6(\pm 37.7)$ oocytes per gram of body mass for each area, respectively. No significant differences were found between the Gulf of Roses and the Ebre 299 Delta either in BF (ANCOVA, $F_{1,12}=0.86$, p=0.377) or in RBF (ANOVA, $F_{1,12}=0.22$, p=0.649).

 As a measure of egg quality, the mean dry mass of hydrated oocytes was estimated at 0.032(\pm 0.011) and 0.036(\pm 0.014) mg per egg for the Gulf of Roses and the Ebre Delta, respectively, 302 which meant there was no significant difference between the two areas (t= -0.8089 , p= 0.426 , n=40).

 The prevalence of atresia (Pa) increased in both areas from the 'Spawning Capable' phase to the 'Regressing' phase of the ovaries. No sign of α-atresia was observed in other development stages, such as the 'Developing' or 'Regenerating' phases (Table 3). The Pa also varied and differed seasonally between sampling areas, i.e. from spring to summer in the Ebre Delta, the Pa decreased while in the Gulf of Roses it increased. On the other hand, the mean relative intensity of atresia (R_{Ia}) increased in relation to the season and the stage of ovarian development as the spawning period advanced.

 The transfer function models fitted to the CPUEs with SSTs as an independent variable explained a considerable amount of the variability in CPUEs (Table 4). The amount of variability explained by these models ranged from 71% for the time series of Sant Carles de la Ràpita to 81% for the time series from Roses. Fig. 5 compares the expected CPUE values (from the transfer function models) with the observed (actual) bluefish CPUE for each time series.

 The transfer functions models showed that CPUE correlated positively with SSTs with a time lag of between 0 and 1 months (significant at t-value>|2|=p<0.05) in almost all time series (Table 4). 319 For example, the transfer function model fitted to bluefish CPUE in Roses $(Y_t;$ output; kg vessel⁻¹) 320 and the corresponding SST $(X_t; input; {}^{\circ}C)$ at lags of 0 and 1 month (time lag 0: t-value=2.31, time 321 lag 1: t-value=3.65) is represented by the equation $Y_t=f(X_t)=0.42\cdot X_{t-0}+0.34\cdot X_{t-1}+a_t$. Thus, the model 322 estimates that an additional 1°C of SST leads to $0.42(\pm 0.06)$ kg vessel⁻¹ added to CPUE of this 323 species during the same month, and $0.34(\pm 0.08)$ kg vessel⁻¹ added to CPUE during the following 324 month, giving a total gain of 0.76 kg vessel⁻¹ for both months. The \mathbb{R}^2 value of the transfer function 325 model was 0.81 (residual standard error of 0.94 kg vessel $^{-1}$). Similar patterns were found in the transfer functions of L'Escala, L'Ametlla de Mar, L'Ampolla and St. Carles de la Ràpita (Table 4).

 Finally, the temperature thresholds analysis applied to the transfer function models indicated that the minimum SST that needs to be reached within each study area in order to enhance bluefish CPUE significantly was around 17-18°C since significant t-values (>|2|, p<0.05) were only observed from this threshold to higher SSTs at each fishing port (Table 5). The AIC value (for each time series) decreased as the SST threshold increased, thereby indicating that the most parsimonious models were those obtained at higher SSTs (Table 5), which in turn supports the idea that bluefish CPUE have been closely linked to the sea surface temperatures.

DISCUSSION

 Our analysis of different reproductive and condition indices shows that it is not only in terms of reproduction that bluefish seem to benefit from actual environmental conditions in the north, but they also appear to be in better condition. Moreover, oocytes of the females from the Gulf of Roses were significantly larger than those of females from the Ebre Delta, suggesting they have higher quality eggs. Apparently, despite the warmer environment that prevails in the Ebre Delta, bluefish were in significantly better condition in the northern study area where the waters are colder. This later may be due to ecological differences between northern and southern areas of the NW Mediterranean. The Gulf of Lions is known to be affected by a permanent cyclonic circulation of surface waters and intense vertical convections which promote intense algal blooms and elevated primary production (Bosc *et al.*, 2004). These later authors estimated a 4-year average annual primary production relatively higher for the Gulf of Lions basin (including the Gulf of Roses) from 180 to 204 gC m⁻² y⁻¹ than estimates for the Balearic basin (including the Ebre Delta area) from 348 153 to 175 gC m⁻² y ⁻¹. Regarding the trophic structure and functioning of these two marine ecosystems, the total biomass (excluding detritus) has been estimated for the Gulf of Lions at 68.9 t 350 km -2 (Bănaru *et al.*, 2013) and for the Ebre Delta at 58.99 t km -2 (Coll *et al.*, 2006). It is noteworthy the total net primary production that differed noticeable between these areas, being 352 considerably higher in the Gulf of Lions, 1042.4 t km⁻² y⁻¹ (Bănaru *et al.*, 2013), than that 353 estimated for the Ebre Delta, 386.68 t km ⁻² y ⁻¹ (Coll *et al.*, 2006). Thus, bluefish inhabiting the more northern area appear to be taking advantage of the environmental conditions found there.

 The results concerning the reproductive biology, such as the oocyte size-frequency distribution and the seasonal relative intensity of atresia, provide evidence that bluefish are multiple batch spawners with asynchronous oocyte development and indeterminate fecundity which is in accordance with previous studies (Conand, 1975; Robillard *et al.*, 2008). Our estimates of batch fecundities for bluefish in the two Mediterranean areas we sampled were very similar to each other, but smaller than those estimated for the bluefish in U.S. East Coast (Robillard *et al.*, 2008) or the West African Coast (Conand, 1975), which ranged 0.2-1.4 and 0.6-1.6 million oocytes, respectively. To date, this study represents the first published work that estimates not only the fecundity, but also the size at maturity of this species in the Mediterranean Sea. It should be noticed 364 that the L_{50} values we obtained for females 37.6 cm TL are much lower than those reported for areas in the Atlantic: 48.0 cm TL in the East Coast of the U.S (Robillard *et al.*, 2008) and 43.0 cm TL in the West African Atlantic Coast (Conand, 1975). It is worth to mention that size at maturity was estimated from relatively similar size ranges (TL): 23.0-97.5 cm (Robillard *et al.*, 2008) and 26.0- 91.1 cm (this study). It has been suggested that maturation in fish populations usually might be closely related to total population abundance over the life of a cohort, with cohorts maturing at an earlier age and smaller size when population size is low (Morgan and Colbourne, 1999). Besides the fact that bluefish mature at smaller sizes in the Mediterranean and also that better fish conditioning was observed in the north would somehow facilitate its capacity for expansion since some studies have shown that fish in better condition may have more surplus energy to devote to reproduction (i.e. an increase number and quality of eggs) which can have a direct impact on spawning biomass and growth of a population (Kjesbu *et al.*, 1991; Rideout *et al.*, 2000; Morgan, 2004).

 Our results support the idea previously presented by Sabatés *et al.* (2012) that in the northern areas of the NW Mediterranean the temperature conditions are gradually becoming more favourable to the bluefish reproduction. Although, the spawning of occurs mainly during the summer season in both of our sampling areas the presence of 'Spawning Capable' females and 'Actively Spawning' males during spring in the Ebre Delta (but not in the Gulf of Roses) and a higher proportion of 'Regressing' females during the summer in the Ebre Delta further supported the thesis that spawning activity starts and declines earlier in the more southern Ebre Delta. In addition, a higher prevalence of atresia was found in females from the Ebre Delta earlier during spring and this was also found to decrease earlier in summer. This indicates that active oocyte resorption takes place earlier in the Ebre Delta than it does in the Gulf of Roses.

 Sea temperature may influence the timing of various life processes such as the spawning date (Kjesbu *et al.*, 2010; Morgan *et al.*, 2013). Reproductively active fish from the Ebre Delta were caught when SSTs were between 18.2 and 27.3°C from May to August, while in the Gulf of Roses they were caught when SSTs were between 17.7 and 23.8°C from July to September. Spawning activity started earlier in the Ebre Delta as SSTs rose over 18.0°C in May, in contrast to the Gulf of Roses where spawning individuals were not landed until early summer (July) when SSTs attained similar temperatures, in this case over 17.7°C. However, despite this two-month lag (from May to July), spawning activity was found to begin in both areas at similar temperature thresholds 394 (≈18.0°C). This indicates that, for both areas, temperature may trigger the initiation of spawning activity. In contrast, Sabatés *et al.* (2012) reported that the threshold temperature for spawning was 21°C. The discrepancies with our study may be related to differences in the approaches used to estimate the timing of spawning. In our study, the timing of spawning was estimated by means of a histological study of the gonads, whereas Sabatés *et al.* (2012) used larvae sampling surveys. Other studies have reported that spawning of bluefish in the mid-Atlantic region occurs at temperatures between 17-24°C (Norcross *et al.*, 1974; Kendall and Walford, 1979), while in the Marmara Sea (Turkey) it occurs from early spring until August at SSTs between 20-26°C (Ceyhan *et al.*, 2007).

 The difference in the timing of spawning activity in the NW Mediterranean coast may be the result of the seasonal migratory behaviour usually exhibited by this species. Although, to date, no information is available regarding the migration of bluefish in the Mediterranean, this species is known to have large distributions and show seasonal migratory behaviour, spending the colder months in warm-water areas and, when the surface temperature reaches a certain value, migrating towards cooler waters where the species spawns once a threshold temperature has been attained (Juanes *et al.*, 1996; Shepherd *et al.*, 2006). We suspect the bluefish from the north move towards the south, where the waters are warmer, but we do not have any clear evidence of this at present. Sabatés *et al.* (2012) suggest that the actual sea warming trends and the changes in the seasonality of the sea water temperature - particularly during spring months - may be causing changes in

 spawning phenology and the timing of the seasonal arrival of bluefish into more northern areas favouring the northward temperature-dependent expansion of this species. Similarly, the round sardinella (*Sardinella aurita*) has expanded its northernmost limit of distribution in the NW Mediterranean (Sabatés *et al.*, 2006).

 The influence of water temperature on bluefish fisheries is revealed in this study using CPUE and transfer functions, and our findings support the results found by Sabatés *et al.* (2012) who used data on landings and sequential *t-*test analysis to detect regime shifts (STARS method). The use of transfer functions in the present study showed that CPUE correlated positively with SST indicating that an increase of temperature in a particular month is followed by an increase of CPUE in that month and in the following month. Therefore, it seems that higher SSTs contribute to increasing bluefish availability in the NW Mediterranean. Fluctuations in fish availability may be related to environmental factors (Smith and Page, 1996), i.e. increased or reduced availability may reflect the temperatures that appear to be avoided or preferred by certain fish species (Swain *et al.*, 2000; Colbourne and Bowering, 2001).

 Bluefish show behavioural thermoregulation, that is to say they are able to avoid or select the right environmental temperature (Olla and Studholme, 1985). However, within certain thermal limits, their distribution may be based on other ecological factors such as food availability (small pelagic fish) (Juanes and Conover, 1994). In this sense, both the Gulf of Roses and the Ebre Delta have been recognized as important spawning grounds for small pelagic fish (García and Palomera, 1996; Olivar *et al.*, 2001; Palomera *et al.*, 2007); however higher densities of anchovy has been observed in the southern Gulf of Lyon than in the Ebre Delta (Bellido *et al.*, 2008). Furthermore, bluefish have also been found in and around sea-cages used in the aquaculture of the seabass (*Dicentrarchus labrax*; Sanchez-Jerez *et al.* 2008). This interaction has been observed in the Gulf of Roses, where bluefish tend to congregate during spring-summer time around the existing sea-cages or even break into the cages to prey on the cultured seabass (sea-cage farm technician, *pers. comm.*). Although, the existence of seabass cages in the Gulf of Roses and the Ebre Delta area may

 thus enhance the bluefish's food supply, it is not quite clear if the increase in bluefish is related to the culture of seabass in the Gulf of Roses because the farm has been active since 20 years ago.

 On the whole, we hypothesize that the expansion of bluefish in the NW Mediterranean may have been facilitated by the physical conditions (nowadays favourable water temperatures) and also the productivity, so that this new habitat therefore may become a more advantageous ecological niche for this species to exploit.

 The increasing abundance of bluefish in the northernmost parts of the western Mediterranean may have important consequences for other coastal species that may be ecologically affected by predation or competition. Bluefish is a top predator that feeds on small pelagic species such as, the anchovy, the sardine, the round sardinella, the gilthead seabream or the Mediterranean horse mackerel (Juanes and Conover, 1994; Juanes *et al.*, 1996; Sánchez-Jerez *et al.*, 2008). Therefore, bearing in mind the trophic level of bluefish and the fact that sea warming is impinging favourably on its biology, the status of bluefish populations needs to be studied and monitored.

 Sea temperatures can actually act as an oceanic barrier between fish populations by creating temperature boundaries in their geographic ranges (Crow *et al.*, 2007). Although the distribution of bluefish coincides with sea surface temperatures of 18–27°C along the subtropical continental shelves of the world (Juanes *et al.*, 1996), it has been suggested by Goodbred and Graves (1996) that shifts in the distribution ranges of bluefish and contact between isolated populations have resulted from historical changes in water temperature. However, although no genetic studies have been carried out yet on the bluefish within our study area, Pardiñas *et al.* (2010) did report insignificant genetic differentiation between eastern Atlantic Ocean (Cadiz and Canary Islands) and Mediterranean Sea (Alicante) populations of this species. We can therefore hypothesize that there is a homogenous bluefish population within the NW Mediterranean, where seasonal migrations take place along a latitudinal gradient in response to changes in water temperature and food supply.

 In summary, this study provides evidence that a marine fish species such as bluefish, which is able to expand or it is forced to shift its distribution range due to sea warming, may take advantage of other environmental conditions - such as better food availability – in new habitats which will contribute to increasing its energy reserves and reproductive potential and therefore contribute to enhancing the population's productivity in these areas.

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LIST OF FIGURES

 Fig. 1 Map of the north-western Mediterranean basin showing the two study areas (Gulf of Roses and Ebre Delta) and the location of fishing ports along the Catalan coast where sampling of *Pomatomus saltatrix* was carried out during the 2010-2011-2012 period. The upper panel shows the 635 mean (\pm std. dev.) monthly variation of sea surface temperatures (SST, \degree C) during the study period (2010-2012) at the Gulf of Roses and the Ebre Delta.

 Fig. 2 Box-whisker plots of the adjusted means of HSI and GSI (standardized based on GLMs for 638 sex and maturity) per sampling area. Interquartile box range (25-75%), mean (+), median (\boxminus), whiskers (represented as Q1 and Q3 deviations from the smallest and largest observation, respectively) are also given.

641 Fig. 3 Seasonal ovarian and testis development stages frequency (percent abundance) and mean $(\pm$ std. dev.) seasonal variation in the GSI of bluefish during the 2010-2012 reproductive cycles in the Gulf of Roses (all years pooled) and the Ebre Delta (2012). Development stages: early developing (EDEV), developing (DEV), spawning capable (SC), actively spawning (AS), regressing (RGS) and regenerating (RGN). Sample number per season (n) is also given in parentheses, n/s denotes periods in which none sample was caught.

 Fig. 4 Oocyte size-frequency distribution (per cent abundance per 20 μm diameter classes) in an actively spawning bluefish female (with hydrated oocytes).

 Fig. 5 Comparisons between the observed (actual) and the expected bluefish CPUE in two fishing ports in the Gulf of Roses: (a) Roses and (b) L'Escala, and three fishing ports in the Ebre Delta: (c) L'Ametlla de Mar, (d) L'Ampolla and (e) Sant Carles de la Ràpita. The expected CPUE values 652 represent the contribution of the sea surface temperature (SST) to the bluefish CPUE (kg vessel⁻¹).

653 **LIST OF TABLES**

654 **Table 1** Analysis of deviance tables for generalized linear models fitted to HSI and GSI using as 655 categorical predictors the stage of maturity and sex. Significance are shown as *** for p<0.001 and 656 $*$ for p<0.05; d.f. = degrees of freedom.

Variable	Source	d.f.	Residual deviance	Explained deviance $(\%)$	p-value
HSI	Null		94.088		
	Sex		4.334	6.9	***
	Maturity		7.162	11.0	***
	Sex*Maturity		በ 539	153	***
	Model	324		36.4	
GSI	Null		2005.451		
	Sex		73.046	6.2	***
	Maturity		771.885	41.2	***
	Sex*Maturity		36.573	32	$**$
	Model	324			

657

658 **Table 2** Summary of oocyte size distributions for each developmental stage in bluefish ovaries. Size 659 ranges are given for each stage. For CA, Vgt-1, Vgt-2, Vgt-3 and GVM oocyte developmental 660 stages, measurements were obtained directly from histological sections, and for hydrated oocytes, 661 previously selected by applying glycerin, image analysis was used.

Oocyte Development Stage	Samples (n)	Sampling Area	Mean Diameter $(\mu m \pm Std. Dev.)$	Diameter Range (μm)	
Pre-vitellogenic					
Cortical alveolar (CA)	200	Ebre Delta	144.1 (± 25.3)	$100 - 200$	
	200	Gulf of Roses	152.9 (± 20.8)		
Vitellogenic (yolked)					
	200	Ebre Delta	$228.9 \ (\pm 24.2)$		
Early Vitellogenesis (Vtg-1)	200	Gulf of Roses	248.6 (± 25.8)	$200 - 300$	
	200	Ebre Delta	334.5 (± 27.6)		
Mid Vitellogenesis (Vtg-2)	200	Gulf of Roses	345.6 (± 29.2)	$300 - 400$	
	200	Ebre Delta	427.9 (± 30.3)	$400 - 500$	
Advanced Vitellogenesis (Vtg-3)	200	Gulf of Roses	438.6 (± 24.0)		
<i>Maturation</i>					
Germinal Vesicle Migration (GVM)	200	Ebre Delta	529.2 (± 24.4)	500-600	
	200	Gulf of Roses	539.9 (± 26.7)		
Hydration	200	Ebre Delta	757.1 (± 56.9)	600-1000	
	200	Gulf of Roses	$763.5 (\pm 50.7)$		

663 **Table 3** Prevalence of atresia (P_a) and mean relative intensity of atresia (R_{Ia}) for each stage of 664 ovarian development for each season at each study area (NA denotes that no sign of α-atresia was 665 observed).

	P_a (%)		Mean R_{Ia} (% \pm std. dev.)		
Variation source	Gulf of Roses	Ebre Delta	Gulf of Roses	Ebre Delta	
Maturity stage					
Developing	NA	NA	NA	NA.	
Spawning Capable	23.7	17.2	$9.9(\pm 5.9)$	$4.4(\pm 1.1)$	
Regressing	100.0	23.1	$96.3(\pm2.0)$	$29.9(\pm 10.3)$	
Regenerating		NΑ			
Season					
Spring	14.3	50.0	$14.3(\pm 0.1)$	$4.8(\pm 0.1)$	
Summer	27.0	15.4	$26.7(\pm 37.1)$	$17.0(\pm 15.5)$	
Autumn	NA.	NA	NA	NΑ	
Winter	ΝA	NA	NA	NΑ	

666

667

668 **Table 4** Results of the ARIMA transfer functions between CPUE and SST. Arithmetic mean and 669 standard deviations of the CPUE are given; RSEU is the residual standard error of each transfer 670 function; RSE is the residual standard error of each coefficient; R^2 is the variance explained. 671 Significant coefficients at a t-value \geq (p < 0.05) are shown in bold.

Port	$CPUE$ (kg vessel -1)		RSEU	\mathbb{R}^2	Time lag	Coeff.	RSE	t-value
	Mean	Std. Dev.	$(kg \text{ vessel}^1)$		(months)	$(kg \text{ vessel}^1)$	$(kg \text{ vessel}^{-1})$	
					$\mathbf{0}$	0.42	0.06	2.31
Roses	1.21	2.15	0.94	0.81		0.34	0.08	3.65
					2	0.03	0.06	0.59
					$\mathbf{0}$	0.38	0.08	2.34
L'Escala	1.79	2.74	1.21	0.80		0.41	0.08	3.80
					2	0.05	0.08	0.60
L'Ametlla					$\bf{0}$	1.93	0.97	2.36
de Mar	28.92	34.76	15.68	0.80		2.27	0.96	2.19
					$\overline{2}$	0.16	0.94	0.17
					$\mathbf{0}$	2.23	0.19	2.32
L'Ampollo	3.29	4.20	1.93	0.79		3.42	0.23	3.54
					2	0.02	0.14	0.02
					$\mathbf{0}$	7.97	1.85	2.30
St. Carles	63.71	53.05	28.62	0.71		3.84	1.95	2.85
de la Ràpita					2	1.16	1.48	0.78

673 **Table 5** Results of the SST threshold analyses using the ARIMA transfer functions between 674 bluefish CPUEs and SSTs at different thresholds $(SST_{13} + ... + SST_{27})$. Significant coefficients 675 from the time lag of '0 months' at a t-value $> |2|$ ($p < 0.05$) are shown in bold. The Akaike's 676 Information Criterion (AIC) is also shown in parentheses.

Threshold	Port							
	Roses	L'Escala	L'Ametlla de Mar	L'Ampolla	St. Carles de la Ràpita			
T_{13}	0.28(2492)	0.78(2580)	0.52(2635)	0.85(2503)	0.61(2751)			
T_{14}	0.39(1971)	1.01(1543)	0.68(1321)	0.91(1450)	1.64(2630)			
T_{15}	0.68(1053)	1.13(1413)	1.08(1310)	0.87(1398)	1.58 (1497)			
T_{16}	1.21(845)	1.14(1424)	1.32(1003)	1.69(928)	1.36 (1399)			
T_{17}	2.45(721)	2.09(1387)	1.95(991)	1.83(812)	2.09(967)			
T_{18}	2.68(760)	2.21(1326)	2.13(945)	3.56(820)	2.07(841)			
T_{19}	2.97(612)	2.62(955)	2.31(819)	4.45(736)	2.16(804)			
T_{20}	3.34(694)	2.54(838)	3.33(809)	4.84(735)	3.91(629)			
T_{21}	3.28(657)	2.93(705)	2.41(795)	3.96(601)	3.93(612)			
T_{22}	3.56(535)	3.12(645)	3.01(663)	3.01(647)	3.75(506)			
T_{23}	3.68(501)	3.55(537)	3.93(661)	3.23(473)	3.83(494)			
T_{24}	3.34(589)	3.11(525)	3.84(607)	3.72(414)	3.09(337)			
T_{25}	4.45(423)	3.17(424)	3.05(541)	3.31(415)	3.25(345)			
T_{26}	4.98 (425)	4.33(416)	3.64(552)	3.48(311)	3.22(234)			
T_{27}	4.51 (422)	4.19(313)	3.99(501)	3.94 (398)	3.77(293)			