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1 **Mediated food and hydrodynamics on the ingestion of microplastics by *Daphnia magna****

2

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6

7 **Abstract**

8

9 There is consensus on the need to study the potential impact microplastics (MP) have on
10 freshwater planktonic organisms. It is not yet fully understood how MP enter the aquatic food
11 web or the effect they have on all the trophic levels. As a result of the potential for MP to
12 accumulate throughout food webs, there is increasing interest in evaluating their fate in a
13 variety of environmental conditions. This study investigated the variability in the ingestion of
14 MP to food ratios and the exposed time of MP to *Daphnia magna* in non-sheared and sheared
15 conditions. The sheared environment provided *Daphnia magna* with the conditions for optimal
16 filtering capacity. Regardless of the ratios of MP concentration to food concentration (MP:Food),
17 the filtration capacity of the *Daphnia magna* was enhanced in the sheared experiments. In both
18 the sheared and non-sheared experiments, filtration capacity decreased when the ratios of MP
19 to food concentration and the exposure times to MP were increased. Mortality was mainly
20 enhanced in the non-sheared conditions at higher MP concentrations and exposure times to
21 MP. No mortality was found in the sheared conditions for the exposure times studied. Therefore,
22 in aquatic systems that undergo constant low sheared conditions, *Daphnia magna* can survive
23 longer when exposed to MP than in calm conditions, provided food concentrations do not limit
24 their capacity to filter.

25

26 **Keywords:** Microplastics, Ingestion, Hydrodynamics, Filtration Capacity, *Daphnia magna*

27 **Declaration of interest:** none.

28 **Highlights:**

29 1) The filtration capacity of *D. magna* is enhanced in sheared flows.

30 2) High MP concentrations lead to a reduction in *D. magna* filtration capacity.

31 3) The amount of exposure time to MP reduces the filtration capacity of *D. magna*.

32

33

34 **1. Introduction**

35

36 Plastic has become virtually indispensable in our daily lives. In 2016, a total of 335 million tons
37 of plastic was produced worldwide (Eerkes-Medrano et al., 2015; Horton et al., 2017; Phuong et
38 al., 2016; Plastics Europe, 2018), leading to an ever-increasing quantity of plastic waste, much
39 of which ends up in marine and freshwater environments (Andrady et al., 2003). Microplastics,
40 MP, are globally recognized as an emerging environmental contaminant (Hurley et al., 2018).
41 Although large plastic items initially attracted most attention, microscopic plastic fragments
42 have also been proven to contaminate sediments in beaches, and estuarine and subtidal areas
43 (Thompson et al., 2004). Based on current trends, a future scenario predicts that forty billion
44 tons of plastic will be produced by 2050 (Zalasiewicz et al., 2016). Many authors have studied
45 the effect MP (<5mm in size) have on marine organisms (Andrady et al., 2011; Beiras et al., 2018;
46 de Sá et al., 2018; Van Cauwenberghe et al., 2015; Wright et al., 2013) and, to a lesser extent,
47 on freshwater organisms (Eerkes-Medrano et al., 2015, Rist et al., 2017; Scherer et al., 2017;
48 Wagner et al., 2014). Microplastics often contain chemicals that were incorporated during
49 manufacture (Fries et al., 2013) or absorbed from the environment (Velzeboer et al., 2014) and
50 which may be toxic to organisms, including humans (Rist et al., 2018), if ingested (Rochman et
51 al., 2013; Tanaka et al., 2013). Moreover, MP ingestion may have physical consequences such
52 as obstructing the gut of the organism, or disrupting feeding and digestion (Canniff and Hoang,
53 2018; Cole et al., 2013; Setälä et al., 2014) and thus affecting the fitness of the organism, i.e.,
54 the reproduction of organisms can be altered by decreasing the oocyte number or sperm
55 velocity and also increasing haemocyte mortality (Besseling et al., 2014; Paul-Pont et al., 2016;
56 Sussarellu et al., 2016).

57

58 Even though the great majority of plastic debris ends up in the oceans (Rochman et al. 2013),
59 freshwater ecosystems are often more exposed to MP because of their greater proximity to

60 densely human-populated areas, along with receiving effluent from wastewater treatment
61 plants (Eerkes-Medrano et al., 2015). However, even sites with low flow or situated further
62 down river have also been found to have high amounts of MP as, although these sites have a
63 low population density, they are exposed to high agricultural use (Kapp and Yeatman, 2018).
64 The low density, shape and size of MP enable them to travel extremely easily through water
65 bodies, often remaining suspended in the pelagic zone.

66

67 One mounting concern about MP is because of their small size. They can enter the aquatic food
68 web at very low levels (e.g. plankton) and later affect higher trophic levels (Besseling et al., 2014;
69 Setälä et al., 2014; Wright et al., 2013). As such, species that play important roles in aquatic
70 environments are of particular concern. For example, *D. magna* is a filter feeder key
71 zooplanktonic species present in many freshwater ecosystems and plays an important role in
72 transferring energy through the different levels of the food webs (Elser et al., 2000). Moreover,
73 as *D. magna* also perform diel vertical migration (Lampert et al., 1993), they potentially
74 contribute to transporting MP further to predators occupying different depths of the water
75 column (Wright et al., 2013).

76

77 As *D. magna* have been reported to ingest particles ranging from nanometres to micrometres
78 in size (Burns, 1968; Pau et al., 2013; Serra et al., 2014; Serra et al., 2018), they are likely to
79 ingest MP. Indeed, in laboratory conditions, *D. magna* have been proven to ingest MP (Canniff
80 and Hoang, 2018; Ogonowski et al., 2016; Rist et al., 2017; Scherer et al., 2017, Wagner et al.,
81 2014). However, MP ingestion has been observed to significantly decrease in the presence of
82 additional particles of natural matter, such as algae or sand (Ogonowski et al., 2016; Rist et al.,
83 2017; Scherer et al., 2017), thus reducing exposure to potentially toxic MP.

84

85 To assess the potential effects MP have on *D. magna*, the present study aims to evaluate the
86 mediation in the MP:food ratio using two new parameters. First, by investigating the role
87 hydrodynamic conditions play on the *D. magna* ingestion rate of MP, and second, by exploring
88 *D. magna* filtration efficiency under differing exposure times to MP. The hypotheses were as
89 follows: 1) the filtration capacity of *D. magna* is higher in sheared conditions compared to
90 quiescent flows, 2) the presence of high MP concentrations leads to a low filtration capacity of
91 *D. magna* and 3) increasing exposed time to MP reduces filtration capacity of *D. magna*.

92

93 **2. Methods**

94

95 **2.1. *Daphnia magna*.**

96 The age of the individuals was chosen based on optimizing the filtering efficiency as a function
97 of the characteristic body length of the *Daphnia* individuals which, in turn, is optimal around a
98 week of life (Wickramarathna et al. 2014).

99

100

101

102

103 *D. magna* were collected from three laboratory cultures that were maintained in 40 L containers
104 at a temperature of 20.0 ± 0.5 °C, and at a natural daylight photoperiod and continuous air
105 supply (to avoid anoxia) for two years at the University of Girona (Spain). Individuals were fed
106 three times a week (Monday, Wednesday and Friday) with a mixture of dry spirulina powder
107 (100% *Spirulina platensis*, KeyPharm, Belgium) and Baker's yeast (*Saccharomyces cerevisiae*,
108 Mondeléz International, Spain). One third of the water from the culture was renewed once every
109 two weeks (Müller et al., 2018; Serra et al., 2018; Serra et al., 2019a; Serra et al., 2019b). Mineral
110 water rich in calcium (constant value of 35.7 mg/L) was used to avoid calcium depletion (Riessen

111 et al., 2012). The individuals used in the experiments were seven days old. The age of the
112 individuals was chosen based on optimizing the filtering efficiency as a function of the
113 characteristic body length of the *Daphnia* individuals (Serra et al., 2018) which, in turn, is
114 maximized in the first two weeks of life (Wickramarathna et al. 2014). To control the age of the
115 individuals, ephippia eggs from the laboratory cultures were hatched and left in a dark box for
116 ten days and then continuously lighted afterwards. The *Daphnia* new-borns were fed with
117 spirulina daily until day 7. By analysing a video recording of 10 *D. magna* with the ImageJ
118 software, the mean body length of the individuals was determined at the start of the
119 experiments (2.0 ± 0.2 mm) and during the evolution of the experiments. During the
120 experiments in both non-sheared, (shear rate of $G=0 \text{ s}^{-1}$ i.e., calm conditions), and sheared, shear
121 rate of $G=2.1 \text{ s}^{-1}$ (Serra et al., 2018), and especially for those with high concentrations of MP, the
122 mean body length of the individuals did not show any significant increase (Table 1). All tests,
123 protocols and analyses with *D. magna* were carried out in accordance with the international
124 'OECD/OCDE Guidelines for the Testing' (OECD/OCDE, 1998) and the 'Protocol for the sampling
125 and laboratory testing of invertebrates' code ML-L-I-2013 (ML-L, 2013) of the Ministerio de
126 Agricultura, Alimentación y Medio Ambiente of the Spanish Government.

127

128 **2.2. Microplastics and feeding particles.**

129

130 Standard spherical polystyrene microparticles with a density of $1.05 \text{ g}\cdot\text{mL}^{-1}$ were provided by
131 SonTek/Xylem Inc (San Diego, USA) and were used for the experiments. A spirulina suspension
132 was used to feed the *Daphnia*. To make the spirulina suspension, 1 g of dry spirulina powder
133 (with a density of $1.153 \text{ g}\cdot\text{mL}^{-1}$) was mixed with 1 L of bottled mineral water for 1 min and left
134 for 1 h so that the bigger spirulina particles settled. The supernatant was used as the spirulina
135 suspension for the experiments at a concentration of 50 mL spirulina suspension per litre of
136 bottled mineral water, for which the final food had a particle volume concentration of 8.0 ± 0.5

137 $\mu\text{L L}^{-1}$ (mass concentration of $8.4 \pm 0.5 \text{ mg L}^{-1}$). Therefore, the initial volume concentration for
138 the 0 % MP:100 % food concentration was $8 \pm 0.5 \mu\text{L L}^{-1}$ and, consequently, the initial volume
139 concentration for the 100% MP:0 % food concentration was chosen to be the same, that is, 8.0
140 $\pm 0.1 \mu\text{L L}^{-1}$ (mass concentration of $9.2 \pm 0.1 \text{ mg L}^{-1}$) of MP at their maximum concentration. The
141 particle concentration of spirulina and MP in the desired particle size range was calculated by
142 integrating the concentration of the particles within the range. Since *D. magna* primarily feed
143 on particles $<30 \mu\text{m}$ in diameter, the volume concentration of particles within the range of 2.5
144 to $30 \mu\text{m}$ was calculated and used as a proxy to evaluate particle removal (Serra et al., 2018).
145 Cladocera ingest particles when their size overlaps those of the organic particles they feed on
146 (Arruda et al., 1983; Gliwicz, 1990). This implies that *D. magna* feed on both MP and algae since
147 *D. magna* individuals lack the ability to differentiate food quality.

148
149 The initial particle volume distributions of both the spirulina and MP presented particles in the
150 *Daphnia* ingestible range ($< 30 \mu\text{m}$) (Fig 1). Spirulina also presented larger particles of diameters
151 $> 100 \mu\text{m}$ and MP presented very small particles diameters $< 5 \mu\text{m}$. Both types of particles
152 presented similar particle distributions in the range between 5 and $100 \mu\text{m}$ (Fig. 1). The volume
153 concentration was used as the key parameter to characterize the concentration of food and MP
154 and has been used extensively in laboratory experiments (Müller et al., 2018; Serra et al., 2018;
155 Serra et al., 2019a and Serra et al., 2019b). The water in both the sheared and non-sheared
156 experiments was changed daily throughout the experiments (i.e., on days 2, 3, 4, 5, 6 and 7).
157 The experiments were conducted at a temperature of $20 \pm 1^\circ\text{C}$ and a photoperiod of 12h:12h
158 (light:dark).

159

160 **2.3. Experimental procedures.**

161

162 The sheared experiments were carried out in a plexiglass container filled with 600 mL of water,
163 while the non-sheared experiments, where the underlying mechanism of particle transport is
164 the sedimentation of particles, were conducted in glass containers filled with 2 L of water and
165 maintained at a constant temperature of 20 ± 0.2 °C. Five different MP:algae ratios were fixed:
166 100% MP:0% food concentration (hereafter 100:0 MP), 75-% MP:25% food suspension
167 (hereafter 75:25 MP), 50% MP:50% food suspension (hereafter 50:50 MP), 25% MP:75% food
168 suspension (hereafter 25:75 MP) and 0%-MP:100-% food suspension (hereafter 0:100 MP). The
169 concentrations of polystyrene MP varied between 2.1 mg L^{-1} (25:75 MP) and 8.4 mg L^{-1} (100:0
170 MP), or between $258 \text{ particles mL}^{-1}$ (25:75 MP) and $1031 \text{ particles mL}^{-1}$ (100:0 MP), thus aligned
171 with the MP particle concentration of 3-3000 particles mL^{-1} used by Scherer et al., (2017), and
172 mass MP concentrations of 0.1 to 1 mg L^{-1} employed by Rist et al. (2017). The ratios were
173 achieved by mixing the spirulina suspension with the MP suspension at the corresponding
174 proportions, achieving final volumes of 0.6 L (sheared experiments) and 2 L (non-sheared
175 experiments). Once the corresponding ratios had been achieved, 30 *D. magna* were carefully
176 added to the sheared experiment and 100 *D. magna* to the non-sheared experiment, thus
177 achieving a final *D. magna* concentration of 50 ind L^{-1} .

178

179 For each MP:algae ratio, three replicates were submitted to a sheared environment
180 characterized by a fluid velocity of 3.13 cm s^{-1} , a shear rate of $G = 2.1 \text{ s}^{-1}$ and a turbulent kinetic
181 energy dissipation of $4.4 \cdot 10^{-6} \text{ m}^2 \cdot \text{s}^{-3}$. Shear was chosen to operate within a low shear flow regime
182 in which filtration was enhanced up to 2.6 times that of a steady flow (Serra et al., 2018). This
183 sheared environment was made possible by using a Couette flow device; as described in Serra
184 et al. (2018). Three replicates were carried out for the non-sheared experiments, in which
185 sedimentation took place. Replicates of the non-sheared conditions were made concurrently,
186 while the three replicates of the sheared experiments were carried out consecutively. All

187 sheared and non-sheared experiments began with the *D. magna* being exposed to the food
188 concentration for 24 h, and then continued for seven days at the five fixed MP:algae ratios.

189

190 For both hydrodynamic conditions (sheared and non-sheared experiments), three replicate
191 control experiments, i.e., experiments without *D. magna*, were performed to quantify the
192 particle sedimentation in the five MP:algae ratio experiments. The experiments lasted for either
193 12 h of measurements or until the steady state was achieved.

194

195 **2.4. Filtration capacity.**

196

197 To quantify particle removal by the *D. magna*, the volume concentration of particles, C_t ,
198 removed from the suspension, within their feeding range (2.5 – 30 μm) (Müller et al., 2018;
199 Serra et al., 2018; Serra et al., 2019a; Serra et al., 2019b) was calculated and its ratio over the
200 initial particle volume concentration, C_0 , was analysed. This ratio was calculated with the
201 following expression:

202

$$203 \frac{C_t}{C_0},$$

204

205 where C_t is the volume concentration in suspension at time t , and C_0 is the initial volume
206 concentration.

207

208 The particle volume concentration was measured with a laser particle size analyser (Lisst-100x,
209 Sequoia Inc.) at working particle diameters between 2.5 and 500 μm (Serra et al., 2001). Pau et
210 al. (2013) showed that spirulina particle ingestion by *D. magna* caused an exponential decrease
211 in suspended particle concentrations in non-sheared experiments carried out in the presence of
212 *D. magna*; so that, after 4 h, the initial concentration of spirulina decreased by $e^{-1} = 0.37$.

213 Henceforth, they fixed 4 h as the characteristic time for analysing the ingestion rate of *D. magna*.
214 Given that Serra and Colomer (2016) and Serra et al. (2018) showed that an optimal particle
215 concentration for *D. magna* development was 0.16 µl/L per individual per day, that same
216 concentration rate was used for the 0:100 food experiments as well as for the 100:0 MP
217 experiments. Therefore, for the 25:75 MP experiments, a volume concentration of 0.04 µl/L of
218 MP per individual per day was used. For the 100:0 MP experiments, the particle number
219 concentration was calculated assuming a particle median diameter of 19.8 µm, which resulted
220 in being 1031 particles mL⁻¹, (i.e., within the range used by other authors (Scherer et al., 2017)),
221 and consequently the number of particles for the 25:75 MP experiments was 258 particles mL⁻¹.
222

223
224 Since the suspended particle volume concentration C_t , decreased exponentially with time, it can
225 be expressed as follows:

226
227
$$C_t = C_0 e^{-kt}$$

228 where C_0 is the suspended particle volume concentration at the beginning of the experiment, t
229 is the exposed time and k is the rate of particle removal by both sedimentation (k_s) and *D. magna*
230 filtration (k_{Dph}), i.e., $k = k_s + k_{Dph}$. Therefore, it is possible to write k as

231
232
$$k = -\frac{1}{t} \ln \left(\frac{C_t}{C_0} \right)$$

233
234 and k_s can be calculated from the control experiment without *D. magna* (where $k_{Dph} = 0$), so that
235 k_{Dph} can be determined for the rest of the experiments. The rate of particle removal by *D. magna*
236 filtration depends on the filtering rate of each *D. magna* individual (F , in mL ind⁻¹ L⁻¹) and the *D.*
237 *magna* concentration (C_{Dph}) according to $k_{Dph} = F \times C_{Dph}$, (Pau et al., 2013), thus the filtration
238 capacity can be obtained from

239

$$240 \quad F = \frac{k_{Dph}}{C_{Dph}}$$

241

242 Therefore, the feeding rates of the *D. magna* were investigated for three parameters: sheared
243 and non-sheared conditions, dependence on ~~the ratios of~~ MP to food concentration ratios, and
244 time of exposure to MP.

245

246 3. Results

247 In the control experiments without *D. magna* and for both the non-sheared and the sheared
248 experiments, an exponential decrease in C_t/C_0 was observed with time regardless of the MP:food
249 ratio (Fig. 2). The C_t/C_0 decreased faster as the percentage of MP increased. After 12 h of
250 experiment, the final ratio of C_t/C_0 was greater for the sheared conditions (Fig. 2b) than for the
251 non-sheared (Fig. 2a). The non-sheared 100:0 MP experiment proved full sedimentation of MP
252 at 6 h (Fig. 2a).

253

254 *D. magna* feed on both food and MP at a rate ~~that was~~ dependent on both the MP:food
255 concentrations and hydrodynamic conditions. As for the control experiments, both the non-
256 sheared (Fig. 3a) and sheared experiments (Fig. 3b) showed that in the presence of *D. magna*,
257 the C_t/C_0 was lower than in the experiments without *D. magna* (Fig. 2), proving the individuals
258 ingested ~~the ingestion of~~ both algae and MP by the individuals. For both 0:100 MP and 100:0
259 MP, the C_t/C_0 for sheared experiments was greater than that in the non-sheared conditions.

260

261 *Daphnia* had higher filtration capacity (F) in the sheared than in the non-sheared experiments
262 (Fig. 4a and 4b), and in both the non-sheared and sheared experiments, *Daphnia* exposed to
263 higher concentrations of MP reduced their filtration capacity. In addition, this effect was

264 intensified at longer exposed times to the MP (Fig. 4c and 4d). In the non-sheared experiments,
265 during the first week of exposure to MP, the 100:0 MP at day 5 and the 75:25 MP and 100:0 MP
266 at day 7 could be considered lethal because the filtration $F_{MP:FOOD}/F_{FOOD}$ could not be measured
267 as all *Daphnia* were found dead. For days 1 to 4, the $F_{MP:FOOD}/F_{FOOD}$ decreased with increasing the
268 MP:Food ratio. In the first week of the non-sheared experiments, the $F_{MP:FOOD}/F_{FOOD}$ decreased
269 with increasing the MP:Food ratio but no lethal MP:Food ratio was found since the
270 $F_{MP:FOOD}/F_{FOOD}$, was greater than zero. For the non-sheared and sheared experiments and at the
271 0% MP, the filtration capacity was constant during the seven days of the experiment (Fig. 4c and
272 4d). For the non-sheared experiments with the presence of MP, the filtration remained nearly
273 constant during the first three days of exposure (Fig. 4c) then decreased afterward for longer
274 exposure times (Fig. 4c). The decrease was greater as the percentage of MP increased (Fig. 4c).
275 Similar results were found for the sheared experiments. However, the filtration remained nearly
276 constant for a longer period, i.e., four days (Fig. 4d).

277

278 4. Discussion

279 4.1. Filtration in sheared and non-sheared conditions.

280

281 The hydrodynamic flow regime was found to be a crucial parameter in determining the
282 performance of *D. magna* because it modified their filtering capacity on a mixture of food and
283 MP. As described by Rist et al. (2017), *Daphnia* ingested both MP and food thus proving their
284 lack of ability to differentiate food quality. A reduction of the particle sedimentation was found
285 in the sheared experiments therefore proving that MP could remain in suspension for longer
286 times. In comparison to stagnant conditions (no shear), the low shear dominated flow regime
287 enhanced filtration efficiency for all the exposure times and MP concentrations.

288

289 Rist et al. (2017) found that when daphniids were exposed to 100 nm and 2 µm polystyrene
290 particles, their ingestion was significantly lower than in the algae-only experiment with the
291 reduction being 20.5 % and 6.8 % for a 1 d exposure time. Our results showed that in the non-
292 sheared conditions, for an exposed time to MP of 4 h (first day), the reduction in filtration varied
293 from 2.2 % to 8.2 % for the ratio concentrations of 25:75 MP and 100:0 MP, respectively. In the
294 sheared conditions with the same particle range, the reduction in filtration was lower and varied
295 from 1.9 % to 2.6 % for the ratio concentrations 25:75 MP and 100:0 MP, respectively.
296 ~~However~~Nevertheless, the ingestion of MP does not imply a toxic effect for the organism (Beiras
297 et al., 2018). Canniff and Hoang (2018) found that the ingestion of polyethylene microbeads by
298 *D. magna* does not endanger their survival and reproduction. However, the combination of MP
299 with pollutants like heavy metals (e.g., Cu, Zn, Ni) increases metal accumulation and toxicity to
300 aquatic organisms (Kim et al., 2017; Brennecke et al., 2016). For example, Khan et al. (2015)
301 found that exposing zebrafish (*Danio rerio*) to Ag-incubated MP reduced the uptake of Ag,
302 together with higher proportions of Ag in the intestines of the zebrafish.

303

304 *D. magna* usually feed on organic (bacteria and algae) and inorganic (sludge) particles that lie
305 within the particle-size range of 1-50 µm (Ebert, 2005; Serra et al., 2018). Nevertheless, they can
306 also ingest MP of different types and sizes. In the gut of daphniids, authors have reported the
307 presence of ingested polystyrene beads of 20 nm and 1 µm (Rosenkranz et al., 2009) and of 100
308 nm and 2 µm (Rist et al., 2017), polystyrene spheres of 1 µm and 10 µm (Scherer et al., 2017),
309 and also fluorescent MP like polymethyl methacrylate of 29.5 µm (Imhof et al., 2013) or
310 polyethylene of 6-75 µm (Canniff and Hoang, 2018). Recently, Jemec et al. (2016) proved that
311 *D. magna* can ingest elongated microplastic textile fibres of 300 µm, ~~and also as well as~~ some
312 very large twisted MP fibres around 1400 µm. Those larger microplastic fibres did not produce
313 any daphnid mortality after 2-two days of treatment in experiments where daphniids were pre-
314 fed with algae. Our results show that daphniids present an inability to differentiate MP from

315 food, as was also found by Imhof et al. (2013), indicating a higher risk of MP accumulation in
316 zooplankton feeders thriving in environments presenting low shear mixing. Indeed, on day 1,
317 after 4 h of exposure to MP, the filtration capacity of *D. magna* in the low sheared environments
318 was 3.48 to 1.94 times the filtration capacity in the non-sheared environment for the ratio
319 concentrations of 100:0 MP and 25:75 MP, respectively. In terms of filtration efficiency, on day
320 1 and ~~so on~~there after, filtration deteriorated more significantly upon increasing the MP
321 concentrations in the non-sheared environment, in comparison to the sheared environment.

322
323 The increased ingestion of MP by daphniids in low shear environments reveals a phenomenon
324 never described before. Low shear flows increase the contact frequency between daphniids and
325 suspended particles, thus enhancing *Daphnia magna* ingestion. That is, since the MP are
326 ubiquitously present in the aquatic ecosystems (Scherer et al., 2017), persistent ingestion of MP
327 by the geographically widespread zooplankter *D. magna* in sheared environments might affect
328 organisms in higher trophic levels since the capture of daphniids is promoted in sheared
329 environments (Kjørboe and Saiz, 1995; Romero et al., 2012). In addition, for the same
330 concentration of MP in sheared environments, filtration capacity is higher since shear promotes
331 the interaction between the daphniids and the MP and food particles. Shear enhances daphnid
332 mobility (Serra et al., 2018) which, in turn, results in optimal fitness, higher survival rates, and
333 better conditions for ingestion. Finally, ~~we-it can~~ be speculated that during the diel vertical
334 migration in the water column of freshwater lakes, schooling of *D. magna* (they migrate up and
335 down in groups), can favour the ingestion of suspended MP by each individual. During the
336 movement of the schooling, the mean turbulent energy dissipation has been estimated to vary
337 between $3.4 \cdot 10^{-6}$ and $1.8 \cdot 10^{-6} \text{ m}^2/\text{s}^3$, depending on the size of the individuals (Wickramarathna
338 et al., 2014) which coincides with the $4.4 \cdot 10^{-6} \text{ m}^2/\text{s}^3$ imposed turbulent energy dissipation in the
339 sheared experiments in which ingestion was promoted. The results of the present study indicate
340 that the daphniids thriving in the sheared environment present higher filtering efficiencies for

341 all days of exposure and for all concentrations of MP, therefore, corroborating the first
342 hypothesis of this study: that the filtration capacity of *D. magna* is higher in sheared conditions
343 compared to quiescent flows.

344

345 **4.2. Filtration in the presence of variable MP to food concentrations.**

346

347 Regardless of the sheared conditions and exposure times, the higher the concentration of MP,
348 the lower the capacity of *D. magna* to filtrate. In terms of ingested mass, in the sheared
349 conditions and during the first 4 h exposure, the ingestion of MP by *D. magna* in the higher 100:0
350 MP mass concentration was 3.42 times higher than that of the 25:75 MP mass concentration.
351 As pointed out by Scherer et al. (2017) and Rist et al. (2017), and presuming non-selective
352 feeding, a high concentration of MP leads to a higher encounter rate, which results in an
353 increased feeding rate. This process is exacerbated under sheared conditions. Indeed, for the
354 first day of exposure, after 4 h of exposure to MP, the filtering efficiency of the daphniids in the
355 experiments with MP was only 49% in the quiescent environment and 86% in the sheared.

356

357 Overall, the relationship between the efficiency of filtering ~~capacity~~ capacities and the MP
358 concentrations is quadratic and in accordance with the literature (DeMott, 1982; Porter et al.
359 1982; Scherer et al., 2017; Serra et al., 2014), indicating that given the inability to distinguish
360 between food and MP, the reduction in food ingested has a significant impact on *D. magna*
361 feeding. Contrary to the findings of Aljaibachi and Callaghan (2017) who reported a significant
362 negative impact of MP uptake in the presence of algae, ~~our~~ the results of this study show that
363 filtration was severely reduced in the presence of high MP concentrations, even when food was
364 present. The difference may be attributed to the MP uptake, since Aljaibachi and Callaghan
365 (2017) fed 24-h starved individuals mass concentrations $8.3 \cdot 10^{-5}$ to $5.3 \cdot 10^{-3}$ times lower than
366 those used in ~~our~~ the experiments in the present study. In addition, when MP were added in

367 suspension, Rist et al. (2017) found that after 24 h of exposure the feeding rates for 100 nm and
368 2 µm Polystyrene plastics were 80% and 93% lower than those for experiments with algae only.
369 In ~~our~~ the experiments in the present study, when MP were added and after 28 h of exposure
370 (day 2), filtration rates for the experiments 75:25 MP, 50:50 MP and 25:75 MP were 0.88, 0.93
371 and 0.98 lower than those for the experiments with algae only, thus proving that the presence
372 of MP concentrations leads to lower filtration capacities of *D. magna* -than filtrations capacities
373 for food only.

374

375 **4.3. Filtration on exposure times to MP.**

376

377 Only a few studies report the effects of MP ingestion by zooplankton for time exposures greater
378 than one day. Jemec et al. (2016) proved that *D. magna* can ingest 300 µm and 1400 µm
379 microplastic textile fibres without producing any mortality after ~~2~~ two days of treatment.
380 ~~Also~~ Likewise, Canniff and Hoang, (2018) found *D. magna* can ingest 63-75 µm plastic microbeads
381 that accumulated in their gut but did not affect mortality. Time exposures of 72 h for
382 *Allorchestes Compressa* to polyethylene MP proved no mortality of individuals along with
383 ingestion of MP in the first 12 h of exposure (Chua et al., 2014). Adult *Daphnia* exposed to MP
384 for 21 d showed mortality after 7 d of exposure in treatments where the concentrations of MP
385 were varied (Aljaibachi and Callaghan, 2018), and Rist et al. (2017) showed that after 21 d of
386 exposure to MP, *D. magna* body burdens of particles (mass/animal) of 100 nm or 2 µm increased
387 as MP concentrations increased (from 0.1 mgL⁻¹ to 1 mgL⁻¹). The results of the present study
388 support the third hypothesis of increasing exposed time of MP reduces the filtration capacity of
389 *D. magna*. The filtration efficiency of daphniids decreased with increasing exposure times for all
390 MP concentrations. In addition, exposure to MP in non-sheared conditions ~~results~~ resulted in
391 the mortality of daphniids after 5 d at MP to food ratios larger than 75:25 MP, while after 7 d

392 the critical ratio ~~is~~was 50:50 MP. No mortality was found when daphniids were exposed to
393 sheared conditions, although at the 100:0 MP ratio, i.e., in an environment with full MP
394 presence, the filtering capacity of daphniids on the 7th day of exposure was reduced to 16% of
395 that for the ratio 0:100 MP, i.e., in an environment without MP. Furthermore, for the 75:25 MP
396 it was reduced to 51% for ~~that of~~ the ratio 0:100 MP. Lower reductions in the filtering efficiencies
397 over time were found for the sheared conditions due to the favourable hydrodynamic conditions
398 *Daphnia* had to filter, given that shear enhances the contact between *Daphnia* and MP.

399

400 5. Conclusions

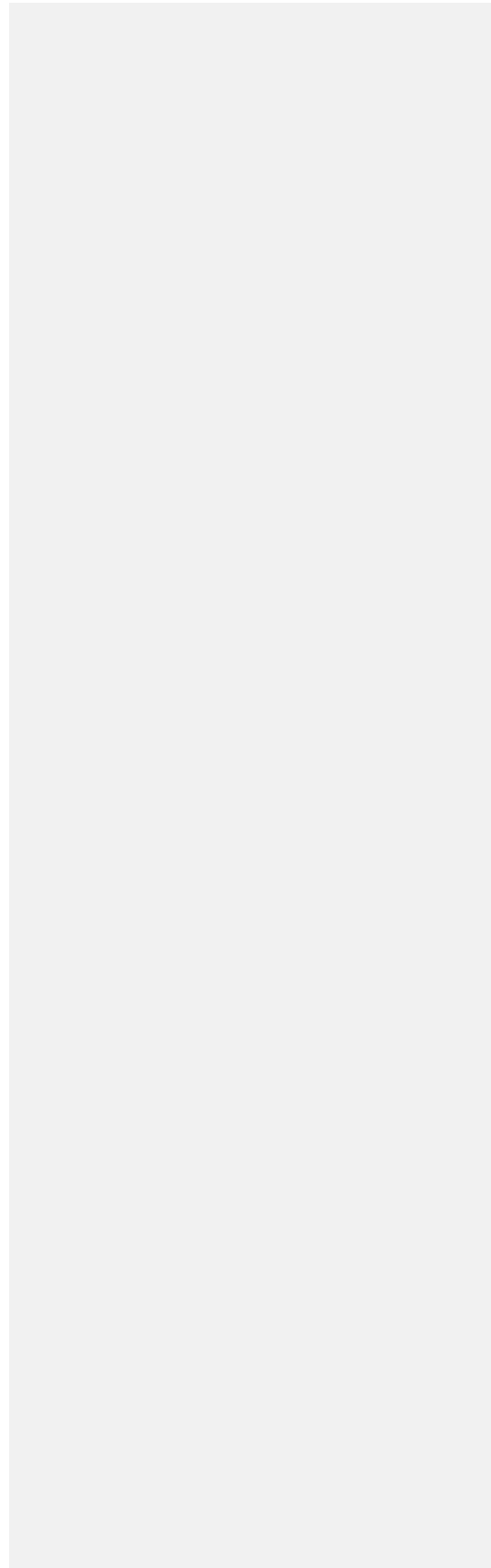
401 Under different controlled characteristics in laboratory tests, the significant phenomena of
402 polystyrene microplastics being ingested by the freshwater zooplanktonic *D. magna* was
403 evaluated. The findings support three hypotheses: that *D. magna* filtration capacity is higher in
404 low shear environments than in quiescent flows, that higher MP concentrations lead to a
405 reduction in *D. magna* filtration capacity, and that at higher exposure times to MP *D. magna*
406 ~~their~~ filtration capacity is reduced. This was especially so for exposure times greater than three
407 days in a quiescent environment with high concentrations of microplastics, and more than four
408 days in sheared environments. Lethal conditions were observed in quiescent flows after 5 d of
409 exposure to MP:food ratios larger than 75:25 MP, while after 7 d the critical ratio was 50:50 MP.

410 Overall, it can be concluded that in the freshwater ecosystems ~~at~~in which MP and *D. magna* are
411 present, the shear produced by mixing can increase the filtration capacity of the individuals,
412 which might affect organisms at higher trophic levels.

413

414 Acknowledgements

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417 Innovation Program 689817.



419 **Captions to Figures.**

420 **Figure 1.** Particle distribution of the initial concentration of spirulina (black dots) and MP (white
421 dots). The limit of *Daphnia magna* particle diameter ingestion range is expressed with a
422 discontinuous line at 30 μm .

423 **Figure 2.** a) Temporal evolution of the ratio C_t/C_0 for the experiments in the non-sheared
424 conditions, for 12 h of experiment. Data corresponds to the control experiments in non-sheared
425 conditions, without *D. magna*, for the experimental MP:food concentrations of 0:100 MP, 25:75
426 MP, 50:50 MP, 75:25 MP and 100:0 MP. The mean and standard deviations for the three
427 replicates are included. b) Temporal evolution of the ratio C_t/C_0 for the experiments in the
428 sheared conditions, for 12 h of experiment. Data corresponds to the control experiments in the
429 sheared condition, without *D. magna*, for the experimental MP:food concentrations of 0:100
430 MP, 25:75 MP, 50:50 MP, 75:25 MP and 100:0 MP. The mean and standard deviations for the
431 three replicates are included.

432 **Figure 3.** a) Temporal evolution of the ratio C_t/C_0 for the experiments in the non-sheared
433 conditions, for 12 h of experiment, for algae only and algae+Daphniids and also for 100:0 MP
434 (microplastics only) and 100:0 MP+ Daphniids. b) Temporal evolution of the ratio C_t/C_0 for the
435 experiments in the sheared conditions, for 12 h of experiment, for algae only and algae+
436 Daphniids and also for 100:0 MP (microplastics only) and 100:0 MP+ Daphniids. The mean and
437 standard deviations for the three replicates (non-sheared experiment) and for the three
438 replicates (sheared experiment) are shown.

439 **Figure 4.** Filtration capacity of daphniids versus concentrations of MP, for different MP:Food
440 ratios, for the times of exposure (days 1, 2, 3, 4, 5 and 7), for both the non-sheared (a) and
441 sheared experiments (b). Filtration capacity efficiency, $F_{\text{MP:FOOD}}/F_{\text{FOOD}}$, as the ratio between the
442 measured filtration capacity of daphniids in the presence of variable concentrations of MP, and
443 the measured filtration capacity in the presence of food, i.e., experiments without the presence
444 of MP, for different MP:Food ratios, for the times of exposure for both the non-sheared (c) and

445 sheared experiments (d). The mean and standard deviations for the three replicates of both
446 (non-sheared experiment) and (sheared experiment) are shown. Results had significant
447 statistical differences (ANOVA test) for experiments with different exposure times (99%
448 confidence) but not within replicas.
449

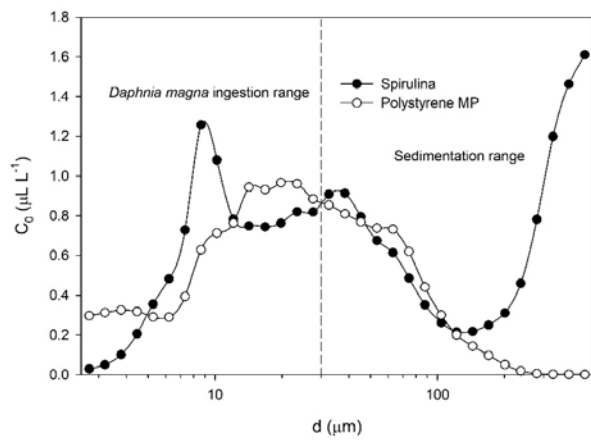


Figure 1

450

451 **Figure 1.** Particle distribution of the initial concentration of spirulina (filled circles) and MP (open
 452 circles). The limit of *Daphnia magna* particle diameter ingestion range is expressed with a
 453 discontinuous line at 30 μm .

454

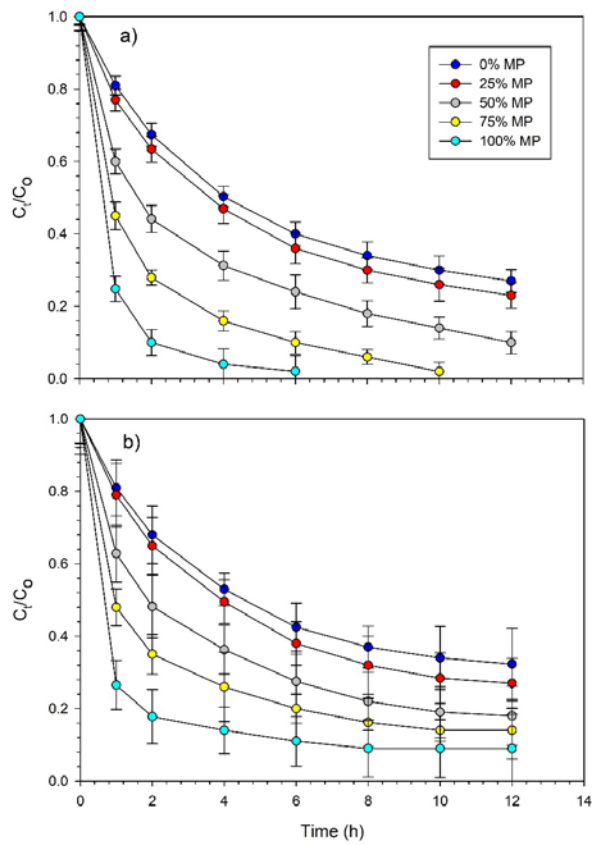


Figure 2

455

456 **Figure 2.** a) Temporal evolution of the ratio C_t/C_0 for the experiments in the non-sheared
 457 conditions, for 12 h of experiment. Data corresponds to the control experiments in non-sheared
 458 conditions, without *D. magna*, for the experimental MP:food concentrations of 0:100 MP, 25:75
 459 MP, 50:50 MP, 75:25 MP and 100:0 MP. The mean and standard deviations for the three
 460 replicates are included. b) Temporal evolution of the ratio C_t/C_0 for the experiments in the

461 sheared conditions, for 12 h of experiment. Data corresponds to the control experiments in the
462 sheared condition, without *D. magna*, for the experimental MP:food concentrations of 0:100
463 MP, 25:75 MP, 50:50 MP, 75:25 MP and 100:0 MP. The mean and standard deviations for the
464 three replicates are included.

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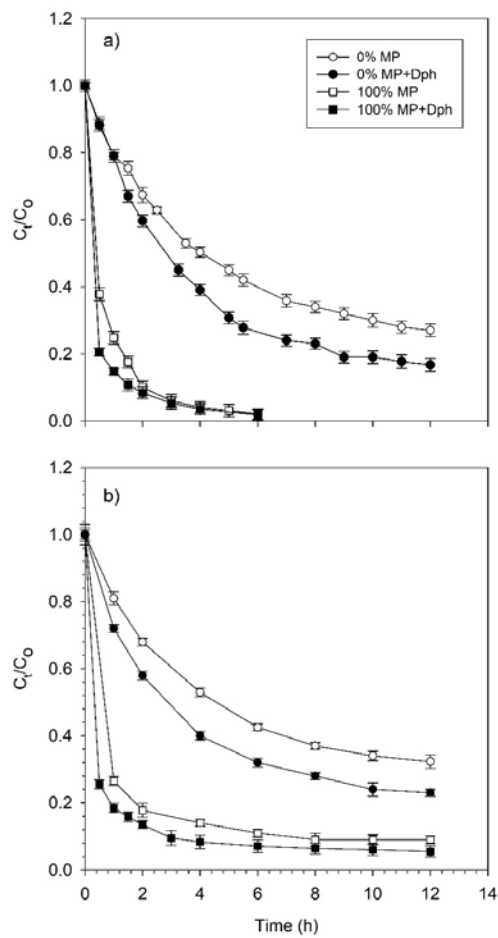


Figure 3

468

469 **Figure 3.** a) Temporal evolution of the ratio C_t/C_0 for the experiments in the non-sheared
 470 conditions, for 12 h of experiment, for algae only and algae+Daphniids and also for 100:0 MP
 471 (microplastics only) and 100:0 MP+ Daphniids. b) Temporal evolution of the ratio C_t/C_0 for the
 472 experiments in the sheared conditions, for 12 h of experiment, for algae only and algae+
 473 Daphniids and also for 100:0 MP (microplastics only) and 100:0 MP+ Daphniids. The mean and

474 standard deviations for the three replicates (non-sheared experiment) and for the three
475 replicates (sheared experiment) are shown.
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477

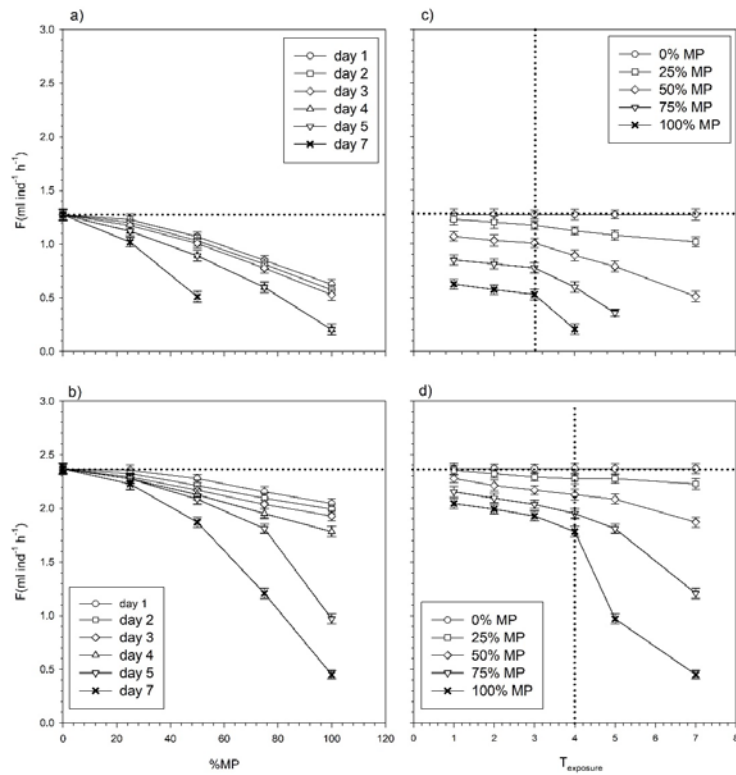


Figure 4

479

480 **Figure 4.** Filtration capacity of daphniids versus concentrations of MP, for different MP:Food
 481 ratios, for the times of exposure (days 1, 2, 3, 4, 5 and 7), for both the non-sheared (a) and
 482 sheared experiments (b). Filtration capacity efficiency, $F_{MP:FOOD}/F_{FOOD}$, as the ratio between the
 483 measured filtration capacity of daphniids in the presence of variable concentrations of MP, and

484 the measured filtration capacity in the presence of food, i.e., experiments without the presence
 485 of MP, for different MP:Food ratios, for the times of exposure for both the non-sheared (c) and
 486 sheared experiments (d). The mean and standard deviations for the three replicates of both
 487 (non-sheared experiment) and (sheared experiment) are shown. Results had significant
 488 statistical differences (ANOVA test) for experiments with different exposure times (99%
 489 confidence) but not within replicas.

490

491 **Table 1.** Sizes of *D. magna* at the end of the 7 d of experiments, in which individuals were
 492 exposed to MP at the experimental MP:food ratio concentrations of 0:100 MP, 25:75 MP, 50:50
 493 MP, 75:25 MP and 100:0 MP, for both sheared and non-sheared conditions. The body sizes of
 494 10 *D. magna* were analysed with the ImageJ software. The mean body length of the individuals,
 495 which was of 2.0 ± 0.2 mm, was determined at the start of the experiments.

496

497	<i>MP:food</i>	<i>0:100 MP</i>	<i>25:75 MP</i>	<i>50:50 MP</i>	<i>75:25 MP</i>	<i>100:0 MP</i>
498	Non-sheared	2.4 ± 0.2 mm	2.2 ± 0.3 mm	2.0 ± 0.2 mm	2.0 ± 0.2 mm	2.0 ± 0.2 mm
499	Sheared	2.4 ± 0.2 mm	2.3 ± 0.2 mm	2.0 ± 0.2 mm	2.0 ± 0.2 mm	2.0 ± 0.2 mm

500

501 **References**

502

503 Aljaibachi, R., Callaghan, A., 2018. Impact of polystyrene microplastics on Daphnia magna in
504 relation to food availability. PeerJ 6:e4601. <http://dx.doi.org/10.7717/peerj.4601>.

505

506 Andrady, A. L., 2003. *Plastics and the environment*; Anthony L. Andrady, Ed.; John Wiley and
507 Sons.

508

509 Andrady, A. L., 2011. Microplastics in the marine environment. *Mar. Pollut. Bull.* 62, 1596–
510 1605. <http://dx.doi.org/10.1016/j.marpolbul.2011.05.030>.

511

512 Arruda, J.A., Marzolf, G.R., Flauk, R.T., 1983. The role of suspended sediments in the nutrition
513 of zooplankton in turbid reservoirs. *Ecology* 64, 1225-1235.

514

515 Beiras, R., Bellas, J., Cachot, J., Cormier, B., Cousin, X., Engwall, M., Gambardella, C., Garaventa,
516 F., Keiter, S., Le Bihanic, F., López-Ibáñez, S., Piazza, V., Rial, D., Tato, T., Vidal-Liñán, L.,
517 2018. Ingestion and contact with polyethylene microplastics does not cause acute
518 toxicity on marine zooplankton. *J. Hazard. Mater.* 360, 452-460.
519 <https://doi.org/10.1016/j.hazmat.2018.07.101>.

520

521 Besseling, E., Wang, B., Lürling, M., Koelmans, A. A., 2014. Nanoplastic affects growth of S.

522 Obliquus and reproduction of D. Magna. *Environ. Sci. Technol.* 48, 12336–12343.

523 <http://dx.doi.org/10.1021/es503001d>.

524

525 Brennecke, D., Duarte, B., Paiva, F., Caçador, I., Canning-Clode, J., 2016. Microplastics as vector
526 for heavy metal contamination from the marine environment. *Estuar. Coast. Shelf. S.* 178, 189-

Formatat: anglès (Regne Unit)

Codi de camp canviat

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527 195. <http://dx.doi.org/10.1016/j.ecss.2015.12.003>.
528
529 Burns, C. W., 1968. The relationship between body size of filter-feeding Cladocera and the
530 maximum size of particle ingested. *Limnol. Oceanogr.* 13, 675–678.
531
532 Canniff, P. M., Hoang, T. C. 2018. Microplastic ingestion by *Daphnia magna* and its
533 enhancement on algal growth. *Sci. Total. Environ.* 633, 500-207.
534 <http://doi.org/10.1016/j.scitotenv.2018.03.176>.
535
536 Cole, M., Lindeque, P., Fileman, E., Halsband, C., Goodhead, R., Moger, J., Galloway, T. S. 2013.
537 Microplastic ingestion by zooplankton. *Environ. Sci. Technol.* 47, 6646-6655.
538 <http://dx.doi.org/10.1021/es400633f>
539
540 Chua, E.M., Shimeta, J., Nugegoda, D., Morrison, P.D., Clarke, B.O., 2014. Assimilation of
541 polybrominated diphenyl ethers from microplastics by the marine amphipod,
542 *Allorchestes Compressa*. *Environ. Sci. Technol.* 48, 8127-8134.
543 <http://dx.doi.org/10.1021/es4057170z>.
544
545 de Sá, L. C., Oliveira, M., Ribeiro, F., Rocha, T. L., Futter, M. N., 2018. Studies of the effects of
546 microplastics on aquatic organisms: what do we know and where should we focus our
547 efforts in the future? *Sci. Total Environ.* 645, 1029–1039.
548 <http://dx.doi.org/10.1016/j.scitotenv.2018.07.207>.
549
550 DeMott, W.R., 1982. Feeding selectivities and relative ingestion rates of *Daphnia* and *Bosmina*.
551 *Limnol. Oceanogr.* 27, 518-527.
552

553 Ebert, D., 2005. Chapter 2 introduction to Daphnia biology. *Ecol. Epidemiol. Parasit. Daphnia* 1-
554 25.

555 Eerkes-Medrano, D., ThomPon, R. C., Aldridge, D. C., 2015. Microplastics in freshwater
556 systems: a review of the emerging threats, identification of knowledge gaps and
557 prioritisation of research needs. *Water Res.* 75, 63–82.
558 <http://dx.doi.org/10.1016/j.watres.2015.02.012>.

559

560 Elser, J. J., Fagan, W. F., Denno, R. F., Dobberfuhl, D. R., Folarin, A., Huberty, A., Interlandi, S.,
561 Kilham, S. S., McCauley, E., Schulz, K. L., Siemann E.H., Sterner, R.W., 2000. Nutritional
562 constraints in terrestrial and freshwater food webs. *Nature* 408 (6812), 578–580.
563 <http://dx.doi.org/10.1038/35046058>.

564

565 Fries, E., Dekiff, J. H., Willmeyer, J., Nuelle, M.-T., Ebert, M., Remy, D., 2013. Identification of
566 polymer types and additives in marine microplastic particles using pyrolysis-GC/MS and
567 scanning electron microscopy. *Environ. Sci. Process. Impacts*, 15, 1949.
568 <http://dx.doi.org/10.1039/C3EM00214D>.

569

570 Gliwicz, Z.M., 1990. Food thresholds and body size in Cladocerans. *Nature* 343, 638-640.

571

572 Horton, A.A., Walton, A., Spurgeon, D.J., Lahive, E., Svendsen, C., 2017. Microplastics in
573 freshwater and terrestrial environments: evaluating the current understanding to
574 identify the knowledge gaps and future research priorities. *Sci. Total Environ.* 586, 127-
575 141. <http://dx.doi.org/10.1016/j.scitotenv.2017.01.190>.

576

577 Hurley, R., Woodward, J.C., Rothwell, J.J., 2018. Microplastic contamination of river beds
578 significantly reduced by catchment-wide flooding. *Nat. Geosci.*, 11, 251-257.
579 <http://dx.doi.org/10.1038/s41561-018-0080-1>.
580
581 Imhof, H.K., Ivleva, N.P., Schmid, J., Niessner, R., Laforsch, C., 2013. Contamination of beach
582 sediments of a subalpine lake with microplastic particles. *Curr. Biol.* 23, R867-R868.
583 <http://dx.doi.org/10.1016/j.cub.2013.09.001>.
584
585 Jemec, A., Horvat, P., Kunej, U., Bele, M., Krzan, A., 2016. Uptake and effects of microplastic
586 textile fibers on freshwater crustacean *Daphnia magna*. *Environ. Pollut.* 219, 201-209.
587 <http://dx.doi.org/10.1016/j.envpol.2016.10.037>.
588
589 Kapp, K.J., Yeatman, E., 2018. Microplastic hotspots in the Snake and Lower Columbia rivers: A
590 journey from the Greater Yellowstone ecosystem to the Pacific Ocean. *Environ. Pollut.*
591 241, 1082-1090. <http://dx.doi.org/10.1016/j.envpol.2018.06.033>.
592
593 Khan, F. R., Syberg, K., Shashoua, Y., Bury, N. R., 2015. Influence of polyethylene microplastic
594 beads on the uptake and localization of silver in zebrafish (*Danio rerio*). *Environ. Pollut.*
595 206, 73-79. <http://dx.doi.org/10.1016/j.envpol.2015.06.009>.
596
597 Kiørboe, T., Saiz, E., 1995. Planktivorous feeding in calm and turbulent environments, with
598 emphasis on copepods. *Mar. Ecol. Prog. Ser.* 122, 135-145.
599
600 Kim, D., Chae, Y., An, Y., 2017. Mixture toxicity of nickel and microplastics with different
601 functional groups on *Daphnia magna*. *Environ. Sci. Technol.* 51, 12852-12858.
602

Formatat: francès (França)

Codi de camp canviat

Formatat: francès (França)

Formatat: francès (França)

603 Lampert, W., 1993. Ultimate causes of diel vertical migration of zooplankton: new evidence for
604 the predator-avoidance hypothesis. [http://hdl.handle.net/11858/00-001M-0000-000F-](http://hdl.handle.net/11858/00-001M-0000-000F-E419-7)
605 [E419-7](http://hdl.handle.net/11858/00-001M-0000-000F-E419-7).
606
607 Müller, M. F., Colomer, J., Serra, T., 2018. Temperature-driven response reversibility and short-
608 term quasi-acclimation of *Daphnia magna*. *PLoS One* 13, e0209705.
609 <https://doi.org/10.1371/journal.pone.0209705>.
610
611 OECD/OCDE. 1998
612
613 Ogonowski, M., Schür, C., Jarsén, Å., Gorokhova, E., 2016. The effects of natural and
614 anthropogenic microparticles on individual fitness in *Daphnia Magna*. *PLoS One* 11,
615 e0155063. <http://dx.doi.org/10.1371/journal.pone.0155063>.
616
617 Pau, C., Serra, T., Colomer, J., Casamitjana, X., Sala, L., Kampf, R., 2013. Filtering capacity of
618 *Daphnia magna* on sludge particles in treated wastewater. *Water Res.* 2013, 47, 181–
619 186. <http://dx.doi.org/10.1016/j.watres.2012.09.047>.
620
621 Paul-Pont, I., Lacroix, C., González-Fernández, C., Hégaret, H., Lambert, C., Le Goïc, N., Frère, L.,
622 Cassone, A., Sussarellu, R., Fabioux, C., Guyomarch, J., Albentosa, M., Huvet, A.,
623 Soudant, P. 2016. Exposure of marine mussels *Mytilus* spp. to polystyrene
624 microplastics: Toxicity and influence of fluoranthene bioaccumulation. *Environ. Pollut.*
625 216, 724-727. <http://dx.doi.org/10.1016/j.envpol.2016.06.039>.
626
627 Plastics Europe. *Plastics-the Facts 2017*; Brussels, 2018.
628

629 Porter, K., Gerritsen, J., Orcutt, Jr. J.D., 1982. The effect of food concentration on swimming
630 patterns, feeding behavior, ingestion, assimilation, and respiration by Daphnia. *Limnol.*
631 *Oceanogr.* 27, 935-949.

632

633 Phuong, N.N, Zalou-Vergnoux, Poirier, L., Kamari, A., Châtel, A., Mouneyrac, Lagarde, F., 2016.
634 Is there any consistency between the microplastics found in the field and those used in
635 laboratory experiments? *Environ. Pollut.* 211, 111-123.
636 <http://dx.doi.org/10.1016/j.envpol.2015.12.035>.

637

638 Protocolo de muestreo y laboratorio de invertebrados bentónicos en lagos. Code : ML-L. 2013

639

640 Riessen, H.P., Linley, R.D., Altshuler, I., Rabus, M., Sollradl, T., Clausen-Schaumann, H.,
641 Laforsch, C., Yan, N.D. 2012. Changes in water chemistry can disable plankton prey
642 defences. *Proc. Natl. Acad. Sci.* 109, 15377-15382. [http://doi.org/10.1038/s41598-019-](http://doi.org/10.1038/s41598-019-40777-2)
643 [40777-2](http://doi.org/10.1038/s41598-019-40777-2).

644

645 Rist, S., Baun, A., Hartmann, N. B., 2017. Ingestion of micro- and nanoplastics in Daphnia
646 magna – quantification of body burdens and assessment of feeding rates and
647 reproduction. *Environ. Pollut.* 228, 398–407.
648 <http://dx.doi.org/10.1016/j.envpol.2017.05.048>.

649

650 Rist, S., Carney Almroth, B., Hartmann, N. B., Karlsson, T. M., 2018. A critical perspective on
651 early communications concerning human health Aspects of microplastics. *Sci. Total*
652 *Environ.* 2018, 626, 720–726. <http://dx.doi.org/10.1016/j.envpol.2018.01.092>.

653

654 Rochman, C. M., Hoh, E., Kurobe, T., Teh, S. J. 2013. Ingested plastic transfers hazardous

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Codi de camp canviat

Formatat: Espanyol (Espanya)

655 chemicals to fish and induces hepatic stress. *Sci. Rep.* 3, 3263.
656 <http://dx.doi.org/10.1038/srep03263>.
657
658 Romero, E., Peters, F., Marrasé, C., 2012. Dynamic forcing of coastal plankton by nutrient
659 imbalances and match-mismatch between nutrients and turbulence. *Mar. Ecol. Prog.*
660 *Ser.* 464, 69– 87. <http://dx.doi.org/10.3354/meps09846>.
661
662 Rosenkranz, P., Chaudhry, Q., Stone, V., Fernandes, T.F., 2009. A comparison of nanoparticle
663 and fine particle uptake by *Daphnia magna*. *Environ. Toxicol. Chem.* 28, 2142-2149.
664
665 Scherer, C., Brennholt, N., Reifferscheid, G., Wagner, M., 2017. Feeding type and development
666 drive the ingestion of microplastics by freshwater invertebrates. *Sci. Rep.* 7, 1–9.
667 <http://dx.doi.org/10.1038/s41598-017-17191-7>.
668
669 Serra, T., Colomer, J., Cristina, X., Vila, X., Arellano, J.B., Casamitjana, X., 2001. Evaluation of a
670 laser in situ scattering instrument for measuring the concentration of phytoplankton,
671 purple sulphur bacteria and suspended inorganic sediments in lakes. *J. Environ. Eng.*
672 127, 1023-1030. [http://dx.doi.org/10.1061/\(ASCE\)0733-9372\(2001\)127:11\(1023\)](http://dx.doi.org/10.1061/(ASCE)0733-9372(2001)127:11(1023)).
673
674 Serra, T., Colomer, J., Pau, C., Marín, M., Sala, L., 2014. Tertiary treatment for wastewater
675 reuse based on the *Daphnia magna* filtration –comparison with conventional tertiary
676 treatments. *Water Sci. Technol.* 70, 705-710. <http://dx.doi.org/10.2166/wst.2014.284>.
677
678 Serra, T., Colomer, J., 2016. The hydraulic retention time on the particle removal efficiency by
679 *Daphnia magna* filtration on treated wastewater. *Int. J. Environ. Sci. Technol.* 13, 1433-
680 1442. <http://dx.doi.org/10.1007/s13762-016-0985-4>.

681

682 Serra, T., Barcelona, A., Soler, M., Colomer, J., 2018. Daphnia magna filtration efficiency and
683 mobility in laminar to turbulent flows. Sci. Total Environ., 621, 626-633.

684 <http://dx.doi.org/10.1016/j.scitotenv.2017.11.264>.

685

686 Serra, T., Colomer, J., Soler, M and Pous, N., 2019a. Daphnia magna filtration, swimming and
687 mortality under ammonium, nitrite, nitrate and phosphate. Sci. Total Environ. 656, 331-

688 337. <https://doi.org/10.1016/j.scitotenv.2018.11.382>.

689

690 Serra, T., Müller, M. F and Colomer, J., 2019b. Functional responses of Daphnia magna to zero-
691 mean turbulence. Sci. Rep-UK 9:3844. <https://doi.org/10.1038/s41598-019-40777-2>.

692

693 Setälä, O., Fleming-Lehtinen, V., Lehtiniemi, M. 2014. Ingestion and transfer of microplastics in
694 the planktonic food web. Environ. Pollut. 185, 77–83.

695 <http://dx.doi.org/10.1016/j.envpol.2013.10.013>.

696

697 Sussarellu, R., Suquet, M., Thomas, Y., Lambert, C., Fabioux, C., Pernet, M. E. J., Le Goïc, N.,
698 Quillien, V., Mingant, C., Epelboin, Y., Corporeau, C., Guyomarch, J., Robbens, J., Paul-
699 Pont, I., Soudant, P., Huvet, A. 2016. Oyster reproduction is affected by exposure to
700 polystyrene microplastics. PNAS. 113, 2430-2435.

701 <http://www.pnas.org/cgi/doi/10.1073/pnas.1519019113>

702

703 Tanaka, K., Takada, H., Yamashita, R., Mizukawa, K., Fukuwaka, M., Watanuki, Y., 2013.

704 Accumulation of plastic-derived chemicals in tissues of seabirds ingesting marine

705 plastics. Mar. Pollut. Bull. 15, 69. <http://dx.doi.org/10.1016/j.marpolbul.2012.12.010>.

706

707 Thompson, R. C., Olse, Y., 2004. Lost at sea: where is all the plastic? *Science* 2004, 304 (5672),
708 838. <http://dx.doi.org/10.1126/science.1094559>.
709
710 Van Cauwenberghe, L., Claessens, M., Vandegehuchte, M. B., Janssen, C. R. 2015. Microplastics
711 are taken up by mussels (*Mytilus edulis*) and lugworms (*Arenicola marina*) living in
712 natural habitats. *Environ. Pollut.* 199, 10-17.
713 <http://dx.doi.org/10.1016/j.envpol.2015.01.008>.
714
715 Velzeboer, I., Kwadijk, C. J. A. F., Koelmans, A. A., 2014. Strong sorption of PCBs to
716 nanoplastics, microplastics, carbon nanotubes, and fullerenes. *Environ. Sci. Technol.* 48,
717 4869–4876. <http://dx.doi.org/10.1021/es405721v>.
718
719 Wagner, M., Scherer, C., Alvarez-Muñoz, D., Brennholt, N., Bourrain, X., Buchinger, S., Fries, E.,
720 Grosbois, C., Klasmeier, J., Marti, T., et al., 2014. Microplastics in freshwater
721 ecosystems: what we know and what we need to know. *Environ. Sci. Eur.* 26, 12.
722 <http://dx.doi.org/10.1186/s12302-014-0012-7>.
723
724 Wickramarathana, L.N., Noss, C., Lorke, A., 2014. Hydrodynamic trails produced by *Daphnia*:
725 size and energetics. *PLoS ONE* 9: e92383.
726 <http://dx.doi.org/10.1371/journal.pone.0092383>.
727
728 Wright, S. L., Thompson, R. C., Galloway, T. S., 2013. The physical impacts of microplastics on
729 marine organisms: a review. *Environ. Pollut.* 178, 483–492.
730 <http://dx.doi.org/10.1016/j.envpol.2013.02.031>.
731
732 Zalasiewicz, J., Waters, C.N., Ivar do Sul, J.A., Corcoran, P.L., Barnosky, A.D., Cearreta, A.,

Formatat: anglès (Regne Unit)

733 Edgeworth, M., Galuszka, A., Jeandel, C., Leinfelder, R., . McNeill, J.R., Steffen, W.,
734 Summerhayes, C., Wagreich, M., Williams, M., Wolfe, A. P., Yonan, Y., 2016. The
735 geological cycle of plastics and their use as a stratigraphic indicator of the
736 Anthropocene. *Anthropocene*, 13, 4-17.
737 <http://dx.doi.org/10.1016/j.ancene.2016.01.002>